

# 丛枝菌根真菌的抑病功能及其应用\*

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**摘要** 土传病害是引发连作障碍的关键因素, 严重制约着我国集约化农业(特别是种植业)的可持续发展。丛枝菌根(Arbuscular mycorrhizal, AM)真菌能够提高宿主抗病性、抑制病原物生长及侵染发病, 是农业生产上防控土传病害的重要有益资源, 探索其抑病机制和应用技术是近年来的研究热点, 但一直缺乏比较系统的理论认识。通过梳理相关研究进展发现, AM真菌主要在宿主根系、根际与植株等3个层面发挥抑病作用, 其中根系防御主要包括AM真菌与病原物竞争生态位和构建机械防御屏障, 根际防御主要包括调节根系分泌物与次生代谢产物及与拮抗菌协同抗病, 而植株防御主要包括促进宿主对养分与水分的吸收和诱导宿主产生系统防御体系, 因而提出AM真菌的“根系-根际-植株三级防御”理论。在应用技术方面, 着重分析通过接种AM真菌或调动土著AM真菌来防控土传病害的技术研究现状, 并对应用中存在的问题及其发展前景进行展望, 一方面可通过建立AM真菌种质资源库或构建转基因工程菌来解决接种有效期及效果稳定性等问题, 另一方面可通过改变施肥模式和调整耕作制度来高效调动土著AM真菌的抑病活性, 旨在为利用AM真菌防控土传病害、促进集约化种植业可持续发展提供理论依据。(图1参157)

**关键词** 土传病害; 连作障碍; 丛枝菌根真菌; 抑病机制; 接种; 施肥

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## The function and potential application of disease suppression by arbuscular mycorrhizal fungi\*

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**Abstract** Soil-borne pathogens are among the key factors preventing continuous cropping, which severely restricts the sustainable development of intensive agriculture (notably plant industries) in China. Arbuscular mycorrhizal fungi (AMF), which can improve the disease resistance of their host plants and inhibit growth and infection by pathogens, are important beneficial resources that can prevent and/or control soil-borne diseases. Although the disease-suppression mechanisms and technological application of AMF have become hot topics for research in recent years, there is a relative lack of a systematic theoretical understanding of these topics. Based on a search and review of the relevant literature, the present study found that AMF act in disease suppression at three main points: in the root, the rhizosphere, and the plant. The root defense pathway of AMF involves niche competition and the construction of a physical defensive barrier against infection. The rhizosphere pathway involves the regulation of root exudates and secondary metabolites and the interaction with other antagonists. The plant pathway involves the improvement of nutrient and water uptake by AMF and the induction of the host's systemic defenses. Therefore, a "three-level defense" theory of AMF is proposed, i.e. the root-rhizosphere-plant defense system. Based on current research on soil-borne disease suppression via inoculation with AMF or stimulating indigenous AMF, the problems and developmental prospects facing the application of AMF were also discussed. One approach that has been suggested to solve

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some of the problems association with the application of AMF, such as the limited term over which treatments remain effective and the low stability of their effects, is the establishment of a germplasm bank of AMF, or the construction of transgenic fungi. On the other hand, stimulating the disease suppression activity of indigenous AMF by changing fertilization patterns and/or the farming system used may also help to solve these problems. These efforts should be made to provide a theoretical basis for the exploiting of AMF for the prevention and control of soil-borne diseases, and the acceleration of the sustainable development of an intensive plant production industry in China.

**Keywords** soil borne disease; continuous cropping obstacle; arbuscular mycorrhizal fungi; disease suppression mechanism; inoculation; fertilization

随着我国农业产业结构的调整,农作物种植趋于规模化、产业化,这在一定程度上解决了农业增产与农民增收问题<sup>[1]</sup>。但由于集约化农业强调土地的高强度利用,且作物种植单一、复种指数高,随着栽培年限延长难免会出现作物生长滞缓、土传病害加剧、产量和品质下降等连作障碍问题<sup>[2-4]</sup>,严重制约农业的可持续发展。连作障碍的成因十分复杂,但主要是土壤、植物、病原物三重因素综合作用的结果,包括土壤理化性质劣化、植物自毒作用、土传病原物激增等<sup>[5-7]</sup>。其中,土传病原物是引起连作障碍极为重要的因素,以真菌为主,也包括部分细菌、线虫和病毒,故主要表现为土壤微生物区系由“细菌型”向“真菌型”转变<sup>[8]</sup>,而真菌化则是土壤健康质量退化的标志<sup>[9]</sup>。连作导致土壤微生物区系发生改变,如芽孢杆菌(*Bacillus*)和假单胞菌(*Pseudomonas*)等有益微生物的数量减少,尖孢镰刀菌(*Fusarium oxysporum*)等病原菌的数量增加<sup>[10-11]</sup>。研究表明,70%的连作障碍是由土传病原物引起的<sup>[12]</sup>,如对连作土壤进行灭菌处理,则能有效缓解再植病害<sup>[13-14]</sup>。然而,在生产实践上对连作土壤进行大规模灭菌存在实施难度和经济制约,且灭菌对土体结构及土壤有益微生物的破坏表明并非一种可持续手段,因而有必要挖掘土壤自身的抑病功能及关键抑病资源。

在土壤生态系统中,有些微生物在抑制土传病原物、促进作物健康生长方面扮演着重要角色,如丛枝菌根(Arbuscular mycorrhizal, AM)真菌正是这样的生防菌<sup>[15]</sup>。AM真菌能与80%以上高等植物建立互惠共生关系<sup>[16]</sup>,能增强植物对非生物胁迫的抗逆性<sup>[17-19]</sup>,而且在防控土传病害方面也有显著的作用效果<sup>[20-21]</sup>。自Safir首次发现接种AM真菌摩西管柄囊霉(*Funneliformis mosseae*)能降低由土棘壳孢(*Pyrenopeziza terrestris*)引起的洋葱根腐病的发病率<sup>[22]</sup>以来,众多研究表明AM真菌能帮助宿主抵御病原物的侵染发病<sup>[23-25]</sup>,如对尖孢镰刀菌和白腐小核菌(*Sclerotium cepivorum*)等均有很好的生防能力<sup>[26]</sup>,对高山红景天根腐病、桉树幼苗青枯病、茄子黄萎病、玉米纹枯病等均有明显的防治效果<sup>[27-29]</sup>。尽管国内外已有大量学者关注AM真菌与土传病原物之间的相互作用,且对其抑病功能展开了长期而丰富的研究,但对其抑病机制和应用技术缺乏系统的整理和认识。因此,本文对AM真菌防控土传病害的作用机制与应用技术研究进展进行梳理总结,以期为利用AM真菌抑制病原物繁殖与侵染宿主、防控土传病害、缓解连作障碍、推动农业可持续发展提供一定参考。

## 1 AM真菌对土传病害的防控作用

AM真菌提高宿主抗病性是一个复杂的综合过程,既可能是局部作用,也可能产生系统性作用<sup>[30]</sup>。综合来看,AM真菌对土传病害的防控作用源自其对植物根系的侵染,进而对宿主植物形成了一个“根系-根际-植株三级防御”体系(图1):首先在根系通过竞争生态位和构建机械防御屏障直接排斥病原物对宿主的侵染,其次在根际通过调节根系分泌物与次生代谢产物及其他拮抗菌产生协同作用来抑制病原物的生长,此外还通过改善宿主的养分与水分状况和诱导植株产生系统防御体系来弥补或减轻病原物侵染对作物造成的危害。因此,在防控土传病害、缓解连作障碍方面表现出巨大的潜力。当然,关于根系、根际与植株的作用位点划分不是绝对的,它们之间是存在交叉或关联的。下面基于“根系-根际-植株三级防御”理论来归纳和讨论AM真菌对土传病害的防控原理。

### 1.1 AM真菌的根系防御作用

1.1.1 AM真菌与土传病原物竞争生态位 引起土传病害的最根本原因是土壤微生物区系失调、病原物激增<sup>[31]</sup>。AM真菌可与病原物竞争生态位,通过抑制病原物繁殖达到防控土传病害的目的<sup>[32]</sup>。生态位竞争主要包括侵染位点和营养能源两个方面。从侵染位点来看,AM真菌侵染宿主根系形成菌根后,其菌丝会迅速占据相应的生态位点,从而减少病原物的侵染位点并降低其数量。例如,在沉香(一种药材)根际接种聚生球囊霉(*Glomus fasciculatum*),根系侵染位点被AM真菌占据,可减轻瓜果腐霉菌(*Pythium aphanidermatum*)的侵染,降低植株病情指数<sup>[33]</sup>;在番茄根际接种摩西管柄囊霉,引起根腐病的寄生疫霉(*Phytophthora parasitica*)在菌根化根系中的入侵位点数明显少于对照<sup>[32]</sup>。研究发现,丛枝结构的发育状况和植物抗病性呈正相关关系,根内根生囊霉(*Rhizophagus irregularis*)侵染后丛枝着生数量与病情指数呈负相关<sup>[34]</sup>。从营养能源来看,AM真菌和病原物的生长都依赖宿主植物提供能量和营养,故存在直接竞争关系<sup>[35]</sup>。当来自宿主根系的光合产物首先被AM真菌利用时,病原物获取的机会无疑会减少,从而限制病原物的生长和繁殖。此外,病原菌及其代谢产物也会抑制AM真菌的生长发育及菌根共生体的形成<sup>[36]</sup>。与之相反,AM真菌还会通过产生拮抗类或营养螯合类物质抑制病原物生长,这则是两者之间具体的竞争策略和竞争手段。

### 1.1.2 AM真菌在根系构建机械防御屏障 AM真菌侵染

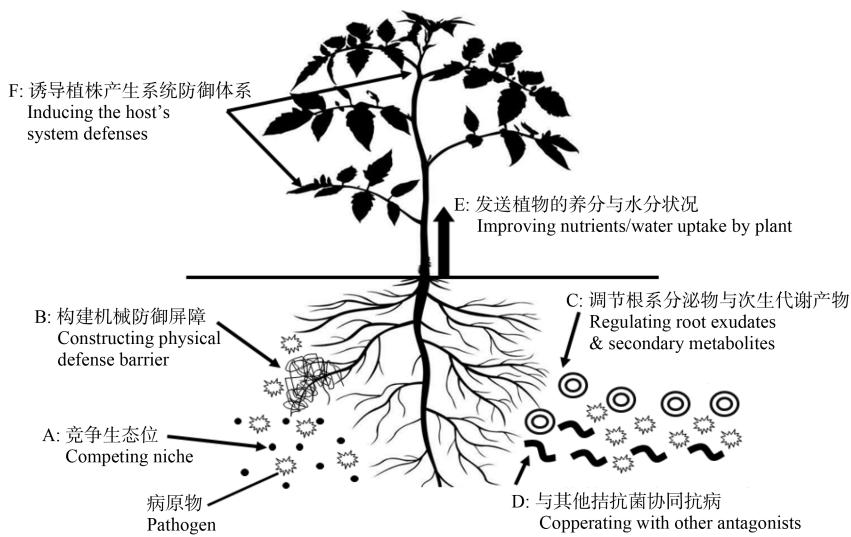


图1 AM真菌对土传病害的“根系-根际-植株三级防御”体系示意图。

Fig. 1 The schematic diagram of “three-level (root-rhizosphere-plant) defense” system of AM fungi to soil-borne diseases.

植物根系形成菌根后,根内与根外蔓延交错的菌丝网建立起一套天然的机械防御屏障,增加病原物侵染植物根系的难度<sup>[37]</sup>。菌根化后根尖表皮会加厚,细胞层数会增加,随着表皮细胞壁物质不断积累,其木质化、硅质化程度不断提高<sup>[38-39]</sup>,且在病原物侵染时会诱导富含羟基脯氨酸的糖蛋白启动快速防御反应使根系细胞壁强度增大,不易被病原物分解的蛋白酶、纤维素酶、半纤维素酶等分泌增强<sup>[40]</sup>,侵染难度随之加大<sup>[41]</sup>。例如,向番茄幼苗单独接种寄生疫霉,根尖分裂组织细胞的分裂活性降低,根尖直径变小,具分裂活性的细胞及细胞核降解或坏死;而同时接种摩西管柄囊霉,根尖分裂组织细胞仍然具有分生能力,根尖变长,直径增大,根皮层相应变厚,对病原菌入侵形成机械屏障,能保护植物根系正常生长<sup>[42]</sup>。利用转移Ri T-DNA胡萝卜根器官双重培养技术研究发现,尖孢镰刀菌菊花专化型(*F. oxysporum* f. sp. *chrysanthemi*)可在非菌根化根系包括维管束中柱等在内的大部分器官中大量繁殖,却被限制在菌根化根系的表皮和外皮层部分<sup>[43]</sup>。

## 1.2 AM真菌的根际防御作用

**1.2.1 AM真菌调节根系分泌物与次生代谢产物** 植物根系因化感作用所分泌的自毒物质会抑制植株防御酶活性<sup>[44-45]</sup>,且可为土传病原物增殖提供所需的营养能源,如番茄、黄瓜、豌豆等根系分泌的苯甲酸、肉桂酸、水杨酸等酚酸类自毒物质会导致病原物侵染作物<sup>[46]</sup>,而外源添加肉桂酸可显著促进尖孢镰刀菌菌丝发育,提高蚕豆枯萎病的病情指数<sup>[47]</sup>。研究发现,AM真菌侵染能减少宿主根系分泌的自毒物质<sup>[48]</sup>,如连作西瓜土壤中酚酸类物质的含量<sup>[49]</sup>。究其原因,根系自毒物质本可直接分泌到土壤中,但菌根形成后会对分泌物进行过滤,其中一部分会被真菌作为营养利用和代谢,所以送达土壤的分泌物会发生很大变化<sup>[50-51]</sup>。同时,菌根化植物的根系分泌物及渗出物(如氨基酸、有机酸等)的数量与质量也会发生变化,进一步改变根际微生物区系<sup>[52]</sup>,如菌根化棉花根际真菌的数量在整个生长期一直低于对照,其

发病率也随之降低<sup>[53]</sup>;接种地表球囊霉(*Glomus versiforme*)的番茄根际细菌数量高于对照,而番茄青枯菌(*Rastonia solanacearum*)的数量却显著低于对照<sup>[54]</sup>。此外,AM真菌能调节植物产生次生代谢产物,如通过提高植株体内苯丙氨酸解氨酶的活性<sup>[40]</sup>来促进和抗病性有关的次生代谢物质(如植保素、生物碱、胼胝质、酚类化合物和类黄酮)的合成<sup>[55]</sup>,在抵御病原菌入侵过程中可降解真菌细胞壁,抑制孢子萌发与菌丝体生长<sup>[56]</sup>;而有的次生代谢产物可产生毒素,抑制病原物的生长繁殖甚至直接杀死病原物<sup>[57-58]</sup>。研究发现,AM真菌既能提高不同连作年限西瓜根内总酚含量,增强西瓜根系抗病性<sup>[49]</sup>;也能诱导西瓜根系产生植保素,显著降低根内和根际土壤中尖孢镰刀菌的数量以及西瓜枯萎病的发病率和病情指数<sup>[59]</sup>;还能促进棉花根系产生大量酚类化合物,提高棉花抵御大丽轮枝菌(*Verticillium dahliae*)的能力<sup>[60]</sup>。另一方面,适宜浓度的次生代谢产物对菌根共生体的建立也能起到促进作用<sup>[61]</sup>。

**1.2.2 AM真菌与其他拮抗菌协同抑制病原物** AM真菌可与土壤有益微生物产生协同抑病作用,如刺激对土传病原物有拮抗作用的微生物(即拮抗菌)的活性,通过吸引木霉(*Trichoderma*)、链霉菌(*Streptomyces*)、粘帚霉(*Gliocladium*)等大量有益拮抗菌到根际定殖,进而消耗大量碳源及其他营养物质<sup>[62-63]</sup>,降低病原物在根际的定殖能力<sup>[64]</sup>。例如,在番茄根际接种摩西管柄囊霉和根内根生囊霉均能增加根际有益拮抗菌数量、抵御病原菌侵染宿主<sup>[65]</sup>;在连作蚕豆根际接种摩西管柄囊霉、扭形球囊霉(*Glomus tortuosum*)和幼套球囊霉(*G. etunicatum*)可不同程度地提高根际细菌的碳源利用能力及多样性指数,同时降低蚕豆枯萎病的病情指数<sup>[66]</sup>。此外,拮抗菌对AM真菌侵染、菌根生长发育及功能发挥也有一定促进作用,如蒙氏假单胞菌(*Pseudomonas monteilii*)能明显促进AM真菌对刺槐的侵染<sup>[67]</sup>,而强壮类芽孢杆菌(*Paenibacillus validus*)可为根内根生囊霉侵染胡萝卜根系提供信号物质<sup>[68]</sup>,这为AM真菌与拮

抗菌协同抑病提供了具体路径。

### 1.3 AM真菌的植株防御作用

**1.3.1 AM真菌改善植物的养分与水分状况** AM真菌能够通过根外菌丝扩大根系吸收范围,有效提高宿主对环境中养分与水分的吸收利用,同时对不同植物间的水分和养分进行再分配<sup>[69]</sup>,从而增加宿主生物量,在一定程度上补偿因病原物侵染而造成的生物量损失和根系功能损伤<sup>[70]</sup>,而健壮的植株也间接提高了抵御病原物侵染的能力<sup>[71-72]</sup>。AM真菌还有助于宿主植物对铁、锌、钴、铜、锰和镍等微量元素的吸收<sup>[73-74]</sup>。例如,发病植株经常表现硼、氮、铁、镁、锌的匮乏<sup>[75]</sup>,而在发生西瓜枯萎病的大田接种AM真菌能促进西瓜对氮、磷、硼和锌等矿质营养的吸收,增加西瓜生物量,降低枯萎病发病率与病情指数<sup>[59]</sup>;给连作番茄接种AM真菌,番茄的叶片中可溶性糖、可溶性蛋白含量显著提高,植株干重随AM真菌侵染率升高而升高<sup>[76]</sup>。研究发现,利用地表球囊霉将黄瓜菌根化育苗后再接种尖孢镰刀菌,壮苗指数较对照提高0.22倍,病情指数较对照降低26.6%<sup>[77]</sup>;民勤绢蒿(*Seriphidium minchiinense*)接种摩西管柄囊霉后叶片可溶性糖与可溶性蛋白含量显著增加,有利于宿主保持较低的渗透势,促进宿主吸收水分,提高其保水能力<sup>[78]</sup>。连作条件下菌根化根系输水能力显著高于普通根系,可以降低植株的萎蔫系数<sup>[79]</sup>。

**1.3.2 AM真菌诱导植物产生系统防御体系** AM真菌能诱导植物更加迅速地产生防御酶类以抵御病原物入侵,其中过氧化氢酶、过氧化物酶和超氧化物歧化酶等可作为植物抗病性的生理生化指标<sup>[80-81]</sup>。研究发现,接种根内根生囊霉可以提高橄榄苗根系超氧化物歧化酶和过氧化物酶活性,从而抑制病原菌大丽轮枝菌的生长<sup>[82-83]</sup>;在葡萄连作土壤接种AM真菌,葡萄叶片与根系超氧化物歧化酶活性均提高,根系活力增强<sup>[84]</sup>。此外,AM真菌侵染作物根系能诱导植株合成内源信号物质,如生长素、细胞分裂素、赤霉素、油菜素内酯、茉莉酸、水杨酸、乙烯、脱落酸等,激活植物的系统防御体系,提高其抗病性<sup>[85-86]</sup>。例如,接种摩西管柄囊霉同时施加外源水杨酸能有效抑制尖镰孢对番茄的侵染,降低病情指数<sup>[87]</sup>;向已感染尖镰孢菌(*F. oxysporum* f.sp. *cucumarinum*)的番茄幼苗根系接种聚生球囊霉和大果球囊霉(*Glomus macrocarpum*),茎叶中茉莉酸的含量提高8倍,且发病指数下降75%-78%<sup>[88]</sup>。AM真菌能提高宿主相关防御基因的转录水平,并通过菌丝桥诱导邻近植株的基因转录<sup>[89]</sup>。病程相关蛋白是植物体内受病原体或其他因子胁迫而诱导表达的一类蛋白,在植物抵御病害方面发挥着重要作用<sup>[90]</sup>。研究发现,丹参接种地表球囊霉后其防御基因PRI和WRKY的表达量显著增加,抗病性显著提高<sup>[91]</sup>;与仅接种早疫病病原菌的对照相比,摩西管柄囊霉侵染的番茄根系中抗病基因的转录水平显著升高,与其有菌丝桥连接的番茄根系中抗性基因的表达量也显著提高<sup>[92]</sup>。通过嫁接试验体系证实,受AM真菌感染诱导的抗病信号甚至还可以从砧木根系远距离传导至地上部接穗的茎叶,进而激发地上部的抗病性<sup>[93]</sup>。

## 2 AM真菌在防控土传病害中的应用

AM真菌防控土传病害的应用技术,可主要分为接种AM

真菌和调动土著AM真菌两大方向。在应用前最好先测定土著AM真菌的活性及数量,若土著AM真菌拥有高数量且活性强,则应该采取保护措施从而更有效地利用土著菌的作用,若土著AM真菌数量少且活性差,则可以考虑引入外来菌种<sup>[94]</sup>。接种AM真菌既可单独接种,也可与其他有益菌(含生防菌)联合接种。调动土著AM真菌主要通过调整施肥模式或改变栽培制度等途径,如通过合理减少氮肥与磷肥投入、增施有机肥以及构建间作系统等来调动土著AM真菌的促生抗病活性<sup>[95-96]</sup>,在防控土传病害上具有较大挖掘潜力。

### 2.1 接种AM真菌

**2.1.1 单接种AM真菌** 研究发现,接种幼套球囊霉显著降低黄瓜枯萎病病情指数和发病率,根际真菌减少,细菌增多,降低了病原菌侵害宿主的机率<sup>[97]</sup>;接种摩西管柄囊霉能降低西瓜枯萎病病情指数、根内和根际土壤尖孢镰刀菌数量<sup>[98]</sup>;接种根内根生囊霉和摩西管柄囊霉可以推迟烤烟青枯病发病时间,降低发病率和病情指数<sup>[99]</sup>;接种地表球囊霉则抑制根际大丽轮枝菌的繁殖,缓解棉花枯萎病发生<sup>[100]</sup>。但是,AM真菌的来源也会影响其接种效应,如在土壤不灭菌条件下接种源自供试土壤本身的AM真菌在增强植物抗病性上显著优于外源的摩西管柄囊霉<sup>[101]</sup>,这与所接种AM真菌的环境适应性有关,也可能是由于土著菌与外源菌之间存在竞争。国内外针对接种单一AM真菌防控土传病害已进行了深入的研究,但目前关于接种AM真菌混合菌剂抑制土传病原物的研究仍不多见。接种混合菌剂往往能更有效地促进宿主吸收营养<sup>[102]</sup>,提高生物量<sup>[103]</sup>,并更有效地抑制土传病原物侵染<sup>[104]</sup>。例如,接种苏格兰管柄囊霉(*Funneliformis caledonii*)与混合菌剂(*Glomus* spp.和*Acaulospora* spp.等)均显著提高AM真菌侵染率,但仅混合菌剂提高黄瓜生物量、降低枯萎病发病率,并达到与未接种尖孢镰刀菌处理相当的产量水平<sup>[105]</sup>;以地表球囊霉和混合菌剂*Glomus* spp.分别对番茄进行菌根化育苗,混合菌剂对番茄青枯病的抑制优于单一菌剂<sup>[106]</sup>。此外,对鹰嘴豆预先接种摩西管柄囊霉、聚生球囊霉和根内根生囊霉组合菌剂有效降低了枯萎菌的密度,病情指数下降高达90%<sup>[107]</sup>。

**2.1.2 AM真菌与其他有益菌双接种** 根际促生菌(Plant growth promoting rhizobacteria, PGPR)是能在根际自由活动且通过固氮、溶磷、拮抗病原物等途径促进植物生长和防控土传病害的有益细菌<sup>[108]</sup>,也能诱导植物产生系统抗性增强抗病能力,目前发现的有假单胞菌、芽孢杆菌、根瘤菌(*Rhizobium*)、农杆菌(*Agrobacterium*)、沙雷氏菌(*Serratia Bizio*)等属<sup>[89]</sup>。PGPR与AM真菌双接种,两者之间可通过协同抑制病原物或提高植物抗病性来共同发挥对土传病害的生防功能<sup>[109-110]</sup>,主要的PGPR有假单胞菌和芽孢杆菌<sup>[111-112]</sup>。研究发现,利用摩西管柄囊霉、根内根生囊霉和地表球囊霉与5个PGPR菌株混合接种,黄瓜根际尖孢镰刀菌数量显著减少、枯萎病发病率有效降低<sup>[113]</sup>。与之相似,摩西管柄囊霉与枯草芽孢杆菌(*Bacillus subtilis*)双接种显著降低秋季草莓冠腐病病情指数、根坏疽程度及恶疫霉合子数<sup>[114]</sup>,地表球囊霉与芽孢杆菌双接种能减轻树番茄根系病害发生程度<sup>[115]</sup>,根内根生囊霉与杰氏假单胞菌(*Pseudomonas jessenii*)或类黄假单胞菌(*P. synxantha*)双接种能提高防御性酶活性,降低

丙二醛含量<sup>[116]</sup>, 苏云金杆菌(*Bacillus thuringiensis*)与5种AM真菌共同接种于齿叶薰衣草也有相同的功效<sup>[117]</sup>. 此外, 马铃薯接种根内根生囊霉和PGPR(*Pseudomonas* sp.)可上调植株体内乙烯响应因子(Ethylene-responsive factor, ERF)的基因表达量, 降低立枯丝核菌对马铃薯的危害<sup>[118]</sup>; 两者双接种可协同增强黄瓜对病原菌的抗性, 且改变病原菌相关防御基因的表达<sup>[119]</sup>. 由此可见, AM真菌与PGPR双接种是通过提高防御酶活性、促进合成抗病信号物质、上调防御基因表达和降低植株体内有毒物质等机制拮抗病原物和提高植物抗病性<sup>[120]</sup>. 因此, 进一步探索和开发AM真菌与PGPR的抗病组合菌剂是十分有意义的.

## 2.2 调动土著AM真菌

**2.2.1 改变栽培制度** 目前国内外利用AM真菌防控土传病害的研究大多关注接种外源AM真菌, 如能定向而高效地调动土著AM真菌, 同样具有巨大的应用价值. 例如, AM真菌的群落组成会因土地利用方式、施肥模式、耕作制度及其他管理措施等的调整而发生演替<sup>[121]</sup>, 也会因宿主植物不同而造成群落结构和侵染水平上的差异<sup>[122-123]</sup>. 研究发现, 免耕能有效促进AM真菌根外菌丝生长并显著提高土壤碱性磷酸酶活性, 4年持续免耕并不会造成土著AM真菌群落结构的退化<sup>[124]</sup>; 免耕主要提高大粒径团聚体中AM真菌的种群丰度, 且可通过提高球囊霉素等促进土壤团聚化<sup>[125]</sup>. 结果表明, 免耕或可通过促进AM真菌的增殖和活性来提高其抑病功能. 传统的单一作物种植模式被认为会降低AM真菌的物种丰富度和侵染力, 因为连作会抑制根际土著AM真菌的活性、减少定殖在作物根系的土著AM真菌数量<sup>[126]</sup>, 而关于间作可提高AM真菌多样性的研究已愈来愈受重视<sup>[127]</sup>, 但参与间作的作物种类也会深刻影响AM真菌群落<sup>[128]</sup>. 研究发现, 间作能有效增加单位面积生物多样性, 使土壤中AM真菌物种丰富度和种群多样性高于单作, 可以直接促进AM真菌的侵染与菌根的形成<sup>[129]</sup>. 例如, 间作韭菜可显著提高番茄根系AM真菌侵染率(比单作高出20%), 且根际尖孢镰刀菌的数量显著降低<sup>[20]</sup>; 间作玉米显著增加土壤中AM真菌的生物量(比马铃薯单作提高18%), 土壤真菌与细菌的生物量比值下降, 促进土壤向高肥效的“细菌型”土壤发展<sup>[130]</sup>. 事实上, 间作本身就是防控土传病害、缓解连作障碍的有效手段, 如间作辣椒处理西瓜枯萎病发病率降低24%<sup>[131]</sup>. 间作能够提高根际土壤养分的有效性、促进植物生长, 提高植株体内防御酶活性、增强根系活力, 降低根际病原菌(*F. oxysporum*)数量、提高植株抗病性<sup>[132]</sup>. 在间作系统中, AM真菌既能与病原物直接发生竞争关系, 又能通过宿主间接发挥抗病作用, 且与间作抑病有一定的叠加效应. 目前有关AM真菌在间作系统抑病过程中所发挥作用的认识十分匮乏, 垂待开展系统研究.

**2.2.2 调整施肥模式** 集约化农业生产中高养分投入是最显著的特征, 但氮肥、磷肥对土著AM真菌侵染率、孢子密度、菌根发育存在抑制作用, 会使宿主植物大量削减或者停止向AM真菌分配光合产物, 导致AM真菌侵染势降低<sup>[133]</sup>, 影响AM真菌发育<sup>[134]</sup>. 研究发现, AM真菌侵染率和孢子密度随磷肥施用量的增加而减少<sup>[135]</sup>, 即使是中等施磷水平也大大降低土壤中AM真菌孢子的数量<sup>[136]</sup>. AM真菌的侵染势在供氮水平较低时随磷含量的增加而降低<sup>[137]</sup>, 在供氮充足情况

下该变化会更加明显<sup>[138]</sup>. 与之相反, 长期缺施磷肥能够保持较高的AM真菌侵染率, 即AM真菌的活性更高, 植物依赖性也更大<sup>[139]</sup>. 施用氮肥也会改变土壤中AM真菌的种类和群落组成<sup>[140-141]</sup>, 但磷肥的影响有时较小<sup>[142]</sup>. 研究发现, 施用大量的氮、磷肥不仅能降低AM真菌的侵染率, 也会改变AM真菌的多样性, 一般球囊霉属的数量会增加, 而其它属的数量会减少<sup>[143]</sup>. 另一方面, 土壤含有丰富的有机质也会对丛枝菌根的形成产生正效应, 增加AM真菌的多样性<sup>[144-145]</sup>, 而适量有机质也有利于提高AM真菌的孢子数<sup>[146]</sup>. 因此, 向贫瘠或缺肥土壤中施有机肥往往可通过增加土壤有益微生物<sup>[147]</sup>和改善土壤质地<sup>[148]</sup>等途径来促进AM真菌菌丝生长和侵染<sup>[149]</sup>, 提高其多样性与功能<sup>[150]</sup>. 例如, 在利用有机肥改良的沙土中AM真菌菌丝的生长加快且侵染率明显提高<sup>[151-152]</sup>, 菌丝、孢囊和总侵染率均与土壤有机质含量呈正相关<sup>[153]</sup>. 通过长期定位试验发现, 潮土中施有机肥能显著增加AM真菌的种群数量、物种丰富度与多样性<sup>[154]</sup>, 但也会降低AM真菌侵染率<sup>[155]</sup>, 这应该与土壤肥力状况、宿主植物类型、AM真菌种类和施肥水平等有关<sup>[144]</sup>. 有益栽试验发现, 当有机肥用量低于2.0 g/kg时, 摩西管柄囊霉的侵染率、孢子密度均高于不施肥处理, 且随有机肥水平的提高而逐渐增高, 但当有机肥用量超过2.0 g/kg时, 各项指标则与对照处理相当或显著低于对照处理<sup>[156]</sup>.

## 3 问题与展望

综上可知, AM真菌在提高植物防御和抵抗病原物侵染的能力、防控土传病害与缓解连作障碍方面表现出巨大的应用前景与生产价值. 尽管目前关于AM真菌与病原物之间的互作研究已经取得了丰富进展, 但要将AM真菌规模应用于集约化农业生产中, 仍有一些亟待解决的理论与技术问题.

(1) AM真菌防控连作土传病害的研究大多关注其抗病效果, 但对防控机制方面的研究不够深入, 究竟起主要作用是哪些机制, 起次要作用的又是哪些过程, 目前还没有认识清楚. 因此, 利用同位素标记、分子生物学和激光共聚焦扫描显微等技术探究植物防御体系与根际微生物群落变化的研究亟待加强, 进而探明AM真菌的抑病机制、对各种信号途径的影响过程以及各个信号物质间的相互联系, 明晰AM真菌对土传病原物的“根系-根际-植株三级防御”体系之间的交叉、关联与协调机制.

(2) 目前关于接种AM真菌的活性有效期及效果稳定性难以保证. AM真菌的积极防效依赖于较高的侵染率<sup>[157]</sup>, 但受宿主基因型、AM真菌种类与病原菌接种时间等因素影响. 未来一方面应开展广泛的大田和保护地试验以及长期定位试验, 深入研究能够使宿主受益最大化的最佳接种条件与管理措施, 建立AM真菌生防效果评价标准; 另一方面应对AM真菌菌种甚至菌株进行甄别筛选, 建立AM真菌种质资源库, 为在不同的宿主-病原物系统中接种相应的AM真菌创造条件. 此外, 还可提取、纯化抗病性AM真菌菌株的抗菌物质(如抗菌蛋白、抗生素等), 克隆、分离能够表达抗菌物质的基因, 通过遗传工程技术, 构建高效、多抗转基因工程菌.

(3) AM真菌是一类严格共生菌, 目前不能单独进行纯

培养, 只能通过侵染宿主植物根系进行扩繁, 使AM菌剂的规模化生产难以完成, 这严重限制了AM真菌菌剂的推广应用。因此, 如何通过改变施肥模式和调整耕作制度等方式来定向调动和充分发挥土著AM真菌的抑病功能, 应是未来需要倍加重视和努力推进的研究方向。

### 参考文献 [References]

- 1 董小艳, 程智慧, 张亮. 百合根系分泌物对4种观赏植物的化感作用[J]. 西北农林科技大学学报(自然科学版), 2008, **36** (9): 113-117 [Dong XY, Cheng ZH, Zhang L. Allelopathy of lily root exudates on some receiver ornamental plants [J]. *J NW A&F Univ (Nat Sci Ed)*, 2008, **36** (9): 113-117]
- 2 黄春生, 熊明. 连作障碍的产生原因及改善途径[J]. 上海蔬菜, 2010 (5): 62-64 [Huang CS, Xiong M. Causes of continuous cropping obstacle and its improvement [J]. *Shanghai Veg*, 2010 (5): 62-64]
- 3 胡海军, 吴亚男, 鄂洋, 高智席, 关晓溪, 张家臣. 设施园艺连作障碍研究进展[J]. 安徽农业科学, 2016 (5): 49-51 [Hu HJ, Wu YN, E Y, Gao ZX, Guan XX, Zhang JC. Research progress of continuous cropping obstacles in protected horticulture [J]. *J Anhui Agric Sci*, 2016 (5): 49-51]
- 4 高群, 孟宪志, 于洪飞. 连作障碍原因分析及防治途径研究[J]. 山东农业科学, 2006 (3): 60-63 [Gao Q, Meng XZ, Yu HF. Reason analysis and control methods of succession cropping obstacle [J]. *Shandong Agric Sci*, 2006 (3): 60-63]
- 5 孙光闻, 陈日远, 刘厚诚. 设施蔬菜连作障碍原因及防治措施[J]. 农业工程学报, 2005, **21** (14): 184-188 [Sun GW, Chen RY, Liu HC. Analysis of obstacle causes of greenhouse vegetable continuous cropping and prevention measures [J]. *Trans Chin Soc Agric Eng*, 2005, **21** (14): 184-188]
- 6 Larkin RP, Griffin TS. Control of soilborne potato diseases using *Brassica* green manures [J]. *Crop Prot*, 2007, **26** (7): 1067-1077
- 7 Jie W, Liu X, Cai B. Diversity of rhizosphere soil arbuscular mycorrhizal fungi in various soybean cultivars under different continuous cropping regimes [J]. *PLoS ONE*, 2013, **8** (8): 72-89
- 8 Wu HS, Yang XN, Fan JQ, Miao WG, Ling N. Suppression of *Fusarium* wilt of watermelon by a bio-organic fertilizer containing combinations of antagonistic microorganisms [J]. *BioControl*, 2009, **54** (2): 287-300
- 9 何飞飞. 设施番茄生产体系的氮素优化管理及其环境效应研究[D]. 北京: 中国农业大学, 2006 [He FF. Study on the optimal management of nitrogen and its environmental effects in the plant tomato production system [D]. Beijing: China Agricultural University, 2006]
- 10 Hua JF, Lin XG, Shen WS, Yin R, Feng YZ. Effects of land use history and inoculation with *Fusarium oxysporum* f. sp. *cucumerinum* on soil nematodes communities [J]. *Helminthologia*, 2008, **45** (4): 204-210
- 11 Shen WS, Lin XG, Gao N, Zhang HY, Yin R, Shi WM, Duan ZQ. Land use intensification affects soil microbial populations, functional diversity and related suppressiveness of cucumber *Fusarium* wilt in China's Yangtze River Delta [J]. *Plant Soil*, 2008, **306** (1-2): 117-127
- 12 Xiao ER, Liang W, He F, Cheng SP, Wu ZB. Performance of the combined SMBR-IVCW system for wastewater treatment [J]. *Desalination*, 2010, **250** (2): 781-786
- 13 Desmond OJ, Edgar CI, Manners JM, Maclean DJ, Schenk PM. Methyl jasmonate induced gene expression in wheat delays symptom development by the crown rot pathogen *Fusarium pseudograminearum* [J]. *Physiol Mol Plant Pathol*, 2006, **67** (3-5): 171-179
- 14 Westphal A, Browne GT, Schneider S. Evidence for biological nature of the grape replant problem in California [J]. *Plant Soil*, 2002, **242** (2): 197-203
- 15 Kennedy AC, Smith KL. Soil microbial diversity and the sustainability of agricultural soils [J]. *Plant Soil*, 1995, **170** (1): 75-86
- 16 Willis A, Rodrigues BF, Harris PJC. The ecology of arbuscular mycorrhizal fungi [J]. *Crit Rev Plant Sci*, 2013, **32** (1): 1-20
- 17 孙吉庆, 刘润进, 李敏. 丛枝菌根真菌提高植物抗逆性的效应及其机制研究进展[J]. 植物生理学报, 2012, **48** (9): 845-852 [Sun JQ, Liu RJ, Li M. Advances in the study of increasing plant stress resistance and mechanisms by arbuscular mycorrhizal fungi [J]. *Plant Physiol Commun*, 2012, **48** (9): 845-852]
- 18 Zhao M, Li M, Liu RJ. Effects of arbuscular mycorrhizae on microbial population and enzyme activity in replant soil used for watermelon production [J]. *Inter J Eng Sci Technol*, 2010, **2** (7): 17-22
- 19 Xun F, Xie B, Liu S, Guo C. Effect of plant growth-promoting bacteria (PGPR) and arbuscular mycorrhizal fungi (AMF) inoculation on oats in saline-alkali soil contaminated by petroleum to enhance phytoremediation [J]. *Environ Sci Pollut Res*, 2015, **22** (1): 598-608
- 20 Hage-Ahmed K, Krammer J, Steinellner S. The intercropping partner affects arbuscular mycorrhizal fungi and *Fusarium oxysporum* f. sp. *lycopersici*, interactions in tomato [J]. *Mycorrhiza*, 2013, **23** (7): 543-549
- 21 Lewandowski TJ, Dunfield KE, Antunes PM. Isolate identity determines plant tolerance to pathogen attack in assembled mycorrhizal communities [J]. *PLoS ONE*, 2013, **8** (4): 613-629
- 22 Sarfir GE. The influence of vesicular arbuscular mycorrhiza on the resistance of onion to *Phyrenochacta terrestris* [D]. Urbana: University of Illinois, 1968
- 23 Schonbeck F, Dehne HW. Damage to mycorrhizal and non-mycorrhizal cotton seedlings by *Thielaviopsis basicola* [J]. *Plant Dis Rep*, 1977, **61** (4): 266-276
- 24 Trotta A, Zavaleta-Mejia E, González-Chavez C, Ferrara-Cerrato R. The use of arbuscular mycorrhizae to control onion white rot (*Sclerotium cepivorum* Berk.) under field conditions [J]. *Mycorrhiza*, 1996, **6** (4): 253-257
- 25 Lingua G, D'Agostino G, Massa N, Antosiano M, Berta G. Mycorrhiza-induced differential response to a yellows disease in tomato [J]. *Mycorrhiza*, 2002, **12** (4): 191-198
- 26 Hao ZP, Christie P, Qin L, Wang CX, Li XL. Control of *Fusarium* wilt of cucumber seedlings by inoculation with an arbuscular mycorrhizal fungus [J]. *J Plant Nutr*, 2005, **28** (11): 1961-1974
- 27 李熙英, 崔东成, 黄世臣. AM菌对高山红景天植株生长及根腐病的影响[J]. 延边大学农学学报, 2013, **35** (2): 115-118 [Li XY, Cui DC, Huang SC. Effects of AM on growth and root rot of *Rhodiola sachalinensis* [J]. *J Agric Sci Yanbian Univ*, 2013, **35** (2): 115-118]
- 28 弓明钦, 陈羽, 王凤珍. AM菌根化的两种桉树苗对青枯病的抗性研究[J]. 林业科学研究, 2004, **17** (4): 441-446 [Gong MQ, Chen Y, Wang FZ. Study on the resistance of two eucalyptus seedlings of AM mycorrhizal on bacterial wilt [J]. *For Res*, 2004, **17** (4): 441-446]
- 29 贺忠群, 李焕秀, 汤浩茹. 丛枝菌根真菌对黄瓜立枯病的影响[J].

- 四川农业大学学报, 2010, **28** (2): 200-204 [He ZQ, Li HX, Tang HR. Effect of arbuscular mycorrhizal fungi on cucumber rhizoctonia rot [J]. *Sichuan Agric Univ*, 2010, **28** (2): 200-204]
- 30 Pozo MJ, Azcónaguilar C. Unraveling mycorrhiza-induced resistance [J]. *Cur Opin Plant Biol*, 2007, **10** (4): 393-398
- 31 Wu F, Wang X, Xue C. Effect of cinnamic acid on soil microbial characteristics in the cucumber rhizosphere [J]. *Eur J Soil Biol*, 2009, **45** (4): 356-362
- 32 Vigo C, Norman JR, Hooker JE. Biocontrol of the pathogen *Phytophthora parasitica* by arbuscular mycorrhizal fungi is a consequence of effects on infection loci [J]. *Plant Pathol*, 2000, **49** (4): 509-514
- 33 Tabin T, Arunachalam A, Shrivastava K, Arunachalam K. Effect of arbuscular mycorrhizal fungi on damping-off disease in *Aquilaria agallocha* Roxb. seedlings [J]. *Trop Ecol*, 2009, **50** (2): 243-248
- 34 Thygesen K, Larsen J, Bodker L. Arbuscular mycorrhizal fungi reduce development of pea root-rot caused by *Aphanomyces euteiches*, using oospores as pathogen inoculum [J]. *Eur J Plant Pathol*, 2004, **110** (4): 411-419
- 35 邢颖, 张莘, 郝志鹏, 赵正雄, 于有志, 陈保冬. 烟草内生菌资源及其应用研究进展[J]. 微生物学通报, 2015, **42** (2): 411-419 [Xing Y, Zhang X, Hao ZP, Zhao ZX, Yu YZ, Chen BD. Biodiversity of endophytes in tobacco plants and their potential application a mini review [J]. *Microbiol China*, 2015, **42** (2): 411-419]
- 36 Liu RJ. Effect of vesicular-arbuscular mycorrhizal fungi on *Verticillium* wilt of cotton [J]. *Mycorrhiza*, 1995, **5** (4): 293-297
- 37 Merrild MP, Ambus P, Rosendahl S, Jakobsen I. Common arbuscular mycorrhizal networks amplify competition for phosphorus between seedlings and established plants [J]. *New Phytol*, 2013, **200** (1): 229-240
- 38 Toljander JF. Interactions between soil bacteria and arbuscular mycorrhizal fungi [D]. Uppsala: Swedish University of Agricultural Sciences, 2006
- 39 徐春美. 丛枝菌根真菌对牡丹根系构型的影响[J]. 山东林业科技, 2014 (4): 12-15 [Xu CM. Effects of arbuscular mycorrhizal fungi on root architecture of peony [J]. *J Shandong For Sci Technol*, 2014 (4): 12-15]
- 40 罗巧玉, 王晓娟, 李媛媛, 林双双, 孙莉. AM真菌在植物病虫害生物防治中的作用机制[J]. 生态学报, 2013, **33** (19): 5997-6005 [Luo QY, Wang XJ, Li YY, Lin SS, Sun L. Mechanism of biological control to plant diseases using arbuscular mycorrhizal fungi [J]. *Acta Ecol Sin*, 2013, **33** (19): 5997-6005]
- 41 崔卫东, 龙宣杞, 补娟, 杨蓉. 黄萎病原菌胁迫对丛枝菌根化棉花幼苗根部防御性酶及超微结构的影响[J]. 新疆农业科学, 2009, **46** (6): 1235-1244 [Cui WD, Long XQ, Bu J, Yang R. Effect of *Verticillium* wilt pathogen stress of cotton seedling root protective enzyme and ultrastructure of arbuscular mycorrhizal fungi [J]. *Xinjiang Agric Sci*, 2009, **46** (6): 1235-1244]
- 42 Fusconi A, Gnavi E, Trotta A, Berta G. Apical meristems of tomato roots and their modifications induced by arbuscular mycorrhizal and soilborne pathogenic fungi [J]. *New Phytol*, 1999, **142** (3): 505-516
- 43 Dassi B, Dumas GE, Gianinazzi S. Different polypeptide profiles from tomato roots following interactions with arbuscular mycorrhizal (*Glomus mosseae*) or pathogenic (*Phytophthora parasitica*) fungi [J]. *Aust J Plant Physiol*, 1995, **22** (1): 87-99
- 44 张晓玲, 潘振刚, 周晓锋, 倪吾钟. 自毒作用与连作障碍[J]. 土壤通报, 2007, **38** (4): 781-784 [Zhang XL, Pan ZG, Zhou XF, Ni WZ. Detoxification and continuous cropping obstacles [J]. *Chin J Soil Sci*, 2007, **38** (4): 781-784]
- 45 王延平, 王华田. 植物根分泌的化感物质及其在土壤中的环境行为[J]. 土壤通报, 2010, **41** (2): 501-507 [Wang YP, Wang HT. Allelochemicals from roots exudation and its environment behavior in soil [J]. *Chin J Soil Sci*, 2010, **41** (2): 501-507]
- 46 李贵菊. 日光温室连作障碍形成的原因及防治措施[J]. 河北农业, 2013 (3): 66-68 [Li GJ. Causes and control measures of continuous cropping obstacle in sunlight greenhouse [J]. *Hebei Agric*, 2013 (3): 66-68]
- 47 董艳, 董坤, 杨智仙, 汤利, 郑毅. 肉桂酸加剧蚕豆枯萎病发生的根际微生物效应[J]. 园艺学报, 2016, **43** (8): 1525-1536 [Dong Y, Dong K, Yang ZX, Tang L, Zheng Y. Microbial impacts of *Fusarium* wilt promoted by cinnamic acid in the rhizosphere of vicia faba [J]. *Acta Horticult Sin*, 2016, **43** (8): 1525-1536]
- 48 Ren SZ, Mallik AU. Selected ectomycorrhizal fungi of black spruce (*Picea mariana*) can detoxify phenolic compounds of *Kalmia angustifolia* [J]. *J Chem Ecol*, 2006, **32** (7): 1473-1489
- 49 马通, 李桂舫, 刘润进, 李敏. AM真菌对连作西瓜根内和根围土壤酚酸类物质和黄酮含量的影响[J]. 北方园艺, 2014 (8): 1-4 [Ma T, Li GF, Liu RJ, Li M. Effects of arbuscular mycorrhizal fungus on the content of phenolic acids and flavonoids in continuous clopping watermelon roots and rhizosphere soils [J]. *Northern Horticul*, 2014 (8): 1-4]
- 50 Nadeem SM, Ahmad M, Zahir ZA, Javaid A, Ashraf M. The role of mycorrhizae and plant growth promoting rhizobacteria (PGPR) in improving crop productivity under stressful environments [J]. *Biotechnol Adv*, 2014, **32** (2): 429-448
- 51 Rambelli A. The rhizosphere of mycorrhizae [M]//Marx DH, Kozlowski TT. Ectomycorrhizae: Their Ecology and Physiology. New York: Academic Press, 1973: 299-349
- 52 宋培玲, 郝丽芬, 李欣州, 张键, 云晓鹏. 丛枝菌根真菌特性及其提高植物抗病性的研究进展[J]. 北方农业学报, 2013 (3): 84-85 [Song PL, Hao LF, Li XZ, Zhang J, Yun XP. The characteristics of arbuscular mycorrhizal fungi and improvement of plant disease resistance [J]. *J Northern Agric*, 2013 (3): 84-85]
- 53 顾向阳, 胡正嘉. VA菌根真菌*Glomus mosseae*对棉花根区微生物量和生物量的影响[J]. 生态学杂志, 1994, **13** (2): 7-11 [Gu XY, Hu ZJ. Effects of VAM fungus *Glomus mosseae* on microbial population and biomass in cotton root zone [J]. *Chin J Ecol*, 1994, **13** (2): 7-11]
- 54 朱红慧, 龙良坤, 羊宋贞, 姚青. AM真菌对青枯菌和根际细菌群落结构的影响[J]. 菌物学报, 2005, **24** (1): 137-142 [Zhu HH, Long LK, Yang SZ, Yao Q. Influence of AM fungus on *Ralstonia solanacearum* population and bacterial community structure in rhizosphere [J]. *Mycosistema*, 2005, **24** (1): 137-142]
- 55 Ortú G, Giuseppe R, Balestrini R, Becker JD, Küster H. Plant genes related to gibberellin biosynthesis and signaling are differentially regulated during the early stages of AM fungal interactions [J]. *Mol Plant*, 2012, **5** (4): 951-954
- 56 Cachinero JM, Cabello F, Jorrín J, Tena M. Induction of different chitinase and beta-1,3-glucanase isoenzymes in sunflower (*Helianthus annuus* L.) seedlings in response to infection by *Plasmopara halstedii* [J].

- Eur J Plant Pathol, 1996, **102** (4): 401-405
- 57 Jaiti F, Meddich A, Hadrami IE. Effectiveness of arbuscular mycorrhizal fungi in the protection of date palm (*Phoenix dactylifera* L.) against bayoud disease [J]. *Physiol Mol Plant Pathol*, 2007, **71** (4-6): 166-173
- 58 Song YY, Cao M, Xie LJ, Liang XT, Zeng RS. Induction of DIMBOA accumulation and systemic defense responses as a mechanism of enhanced resistance of mycorrhizal corn (*Zea mays* L.) to sheath blight [J]. *Mycorrhiza*, 2011, **21** (8): 721-731
- 59 李敏, 刘润进, 李晓林. 大田条件下丛枝菌根真菌对西瓜生长和枯萎病的影响[J]. 植物病理学报, 2004, **34** (5): 472-473 [Li M, Liu RJ, Li XL. Influences of arbuscular mycorrhizal fungi on growth and *Fusarium* wilt disease of watermelon in field [J]. *Acta Phytopathol Sin*, 2004, **34** (5): 472-473]
- 60 补娟, 崔卫东, 罗明, 龙宣杞, 杨蓉. 接种丛枝菌根真菌对棉花生长和黄萎病的影响[J]. 新疆农业科学, 2009, **46** (3): 549-555 [Bu J, Cui WD, Luo M, Long XQ, Yang R. The effect of inoculation with AMF inoculum on growth and *Verticillium* wilt of cotton [J]. *Xinjiang Agric Sci*, 2009, **46** (3): 549-555]
- 61 Yao MK, Desilets H, Charles MT, Boulanger R, Tweddell RJ. Effect of mycorrhization on the accumulation of rishitin and solavetivone in potato plantlets challenged with *Rhizoctonia solani* [J]. *Mycorrhiza*, 2003, **13** (6): 333-336
- 62 Bodker L, Kjoller R, Rosendahl S. Effect of phosphate and the arbuscular mycorrhizal fungus *Glomus intraradices*, on disease severity of root rot of peas (*Pisum sativum*) caused by *Aphanomyces euteiches*. *Mycorrhiza*, 1998, **8** (3): 169-174
- 63 Kasimandri RS, Smith SE, Smith FA, Scott ES. Influence of the mycorrhizal fungus, *Glomus coronatum*, and soil phosphorus on infection and disease caused by binucleate *Rhizoctonia* and *Rhizoctonia solani* on mung bean (*Vigna radiata*) [J]. *Plant Soil*, 2002, **238** (2): 235-244
- 64 刘润进, 陈应龙. 菌根学[M]. 北京: 科学出版社, 2007 [Liu RJ, Chen YL. Mycorrhizology [M]. Beijing: Science Press, 2007]
- 65 Lópezráez JA, Verhage A, Fernández I, García JM, Azcón-Aguilar C. Hormonal and transcriptional profiles highlight common and differential host responses to arbuscular mycorrhizal fungi and the regulation of the oxylipin pathway [J]. *J Exp Bot*, 2010, **61** (10): 2589-2601
- 66 董艳, 董坤, 杨智仙, 汤利, 郑毅. AM真菌控制蚕豆枯萎病发生的国际微生物效应[J]. 应用生态学报, 2016, **27** (12): 4029-4038 [Dong Y, Dong K, Yang ZX, Tang L, Zheng Y. Rhizosphere microbial impacts of alleviating faba bean *Fusarium* wilt with inoculating AM fungi [J]. *Chin J Appl Ecol*, 2016, **27** (12): 4029-4038]
- 67 Duponnois R, Plenchette C. A mycorrhiza helper bacterium enhances ectomycorrhizal and endomycorrhizal symbiosis of Australian *Acacia* species [J]. *Mycorrhiza*, 2003, **13** (2): 85-91
- 68 Hildebrandt U, Ouziad F, Marner FJ, Bothe H. The bacterium *Paenibacillus validus*, stimulates growth of the arbuscular mycorrhizal fungus *Glomus intraradices*, up to the formation of fertile spores [J]. *FEMS Microbiol Lett*, 2006, **254** (2): 258-267
- 69 Smith SE, Jakobsen I, Grønlund M, Smith FA. Roles of arbuscular mycorrhizas in plant phosphorus nutrition: interactions between pathways of phosphorus uptake in arbuscular mycorrhizal roots have important implications for understanding and manipulating plant phosphorus acquisition [J]. *Plant Physiol*, 2011, **156** (3): 1050-1057
- 70 Harrier LA, Waston CA. The potential role of arbuscular mycorrhizal (AM) fungi in the bioprotection of plants against soil-borne pathogens in organic and/or other sustainable farming systems [J]. *Pest Manage Sci*, 2004, **60** (2): 149-157
- 71 Hodge A, Diaz SM. Substantial nitrogen acquisition by arbuscular mycorrhizal fungi from organic material has implications for N cycling [J]. *PNAS*, 2010, **107** (31): 13754-13759
- 72 Laparra J, Malbreil M, Letisse F, Portais JC, Roux C. Combining metabolomics and gene expression analysis reveals that propionyl- and butyryl-carnitines are involved in late stages of arbuscular mycorrhizal symbiosis [J]. *Mol Plant*, 2014, **7** (3): 554-566
- 73 Marschner H, Dell B. Nutrient-Uptake in mycorrhizal symbiosis [J]. *Plant Soil*, 1994, **159** (1): 89-102
- 74 Cabral L, Giachini AJ, Siqueira JO. Arbuscular mycorrhizal fungi in phytoremediation of contaminated areas by trace elements: mechanisms and major benefits of their applications [J]. *World J Microb Biot*, 2015, **31** (11): 1655-1664
- 75 高萍, 李芳, 郭艳娥, 段廷玉. 丛枝菌根真菌和根瘤菌防控植物真菌病害的研究进展[J]. 草地学报, 2017, **25** (2): 236-242 [Gao P, Li F, Guo YE, Duan TY. Advances in AM fungi and rhizobium to control plant fungal diseases [J]. *Acta Agrestia Sin*, 2017, **25** (2): 236-242]
- 76 贺忠群, 贺超兴, 张志斌, 邹志荣, 王怀松. 不同丛枝菌根真菌对番茄生长及相关生理因素的影响[J]. 沈阳农业大学学报, 2006, **37** (3): 308-312 [He ZQ, He CX, Zhang ZB, Zou ZR, Wang HS. Physiological study of tomato growth effects induced by different arbuscular mycorrhizal fungus (AMF) strains [J]. *J Shenyang Agric Univ*, 2006, **37** (3): 308-312]
- 77 王倡宪, 李晓林, 宋福强, 王贵强, 李北齐. 两种丛枝菌根真菌对黄瓜苗期枯萎病的防效及根系抗病相关酶活性的影响[J]. 中国生态农业学报, 2012, **20** (1): 53-57 [Wang CX, Li XL, Song FQ, Wang GQ, Li BQ. Effects of arbuscular mycorrhizal fungi on *fusarium* wilt and disease resistance-related enzyme activity in cucumber seedling root [J]. *Chin J Eco-Agric*, 2012, **20** (1): 53-57]
- 78 贺学礼, 高露, 赵丽莉. 水分胁迫下丛枝菌根AM真菌对民勤绢蒿生长与抗旱性的影响[J]. 生态学报, 2011, **31** (4): 1029-1037 [He XL, Gao L, Zhao LL. Effects of AM fungi on the growth and drought resistance of *Seriphidium minchunense* under water stress [J]. *Acta Ecol Sin*, 2011, **31** (4): 1029-1037]
- 79 王红新, 李富平, 国巧真. AM真菌生长发育影响因素及其对植物作用的研究[J]. 中国土壤与肥料, 2006 (1): 52-56 [Wang HX, Li FP, Guo QZ. The growth influence factor of AM and the function of it for the plants [J]. *Soil Fert Sci Chin*, 2006 (1): 52-56]
- 80 Prasad TK. Role of catalase in inducing chilling tolerance in pre-emergent maize seedlings [J]. *Plant Physiol*, 1997, **114** (4): 1369-1376
- 81 Gechev T, Willekens H, Montagu MV, Inzé D, Van Camp W. Different responses of tobacco antioxidant enzymes to light and chilling stress [J]. *J Plant Physiol*, 2003, **160** (5): 509-515
- 82 贺学礼, 刘嫏, 安秀娟, 赵丽莉. 水分胁迫下AM真菌对柠条锦鸡儿(*Caragana korshinskii*)生长和抗旱性的影响[J]. 生态学报, 2009, **29** (1): 47-52 [He XL, Liu W, An XJ, Zhao LL. Effects of AM fungi on the growth and drought resistance of *Caragana korshinskii* under water stress conditions [J]. *Acta Ecol Sin*, 2009, **29** (1): 47-52]

- 83 Espinosa F, Garrido I, Ortega A, Casimiro I, Álvarez-Tinaut MC. Redox activities and ROS, NO and phenylpropanoids production by axenically cultured intact olive seedling roots after interaction with a mycorrhizal or a pathogenic fungus [J]. *PLoS ONE*, 2014, **9** (6): e100132
- 84 李坤. 葡萄连作障碍机理及调控途径的研究[D]. 沈阳: 沈阳农业大学, 2010 [Li K. Study on the mechanism and regulation of continuous cropping obstacle in grape [D]. Shenyang: Shenyang Agricultural University, 2010]
- 85 Hause B, Mrosk C, Isayenkova S, Strack D. Jasmonates in arbuscular mycorrhizal interactions [J]. *Phytochemistry*, 2007, **38** (23): 101-110
- 86 Fiorilli V, Catoni M, Francia D, Cardinale F, Lanfranco L. The arbuscular mycorrhizal symbiosis reduces disease severity in tomato plants infected by *Botrytis cinerea* [J]. *J Plant Pathol*, 2011, **93** (1): 237-242
- 87 Ei-Khalla SM. Induction and modulation of resistance in tomato plants against *Fusarium* wilt disease by bioagent fungi (arbuscular mycorrhiza) and/or hormonal elicitors (jasmonic acid & salicylic acid): I-changes in growth, some metabolic activities and endogenous hormones related to defence mechanism [J]. *Aus J Basic Appl Sci*, 2007, **1** (4): 691-705
- 88 Rupam K. Induced resistance in mycorrhizal tomato is correlated to concentration of jasmonic acid [J]. *Online J Biol Sci*, 2008, **8** (3): 49-56
- 89 Ismail Y, Hijri M. Arbuscular mycorrhization with glomus irregulare induces expression of potato PR homologous genes in response to infection by *Fusarium sambucinum* [J]. *Funct Plant Biol*, 2012, **39** (3): 236-245
- 90 张玉, 杨爱国, 冯全福. 植物病程相关蛋白及其在烟草中的研究进展[J]. 生物技术通报, 2012 (5): 20-24 [Zhang Y, Yang AG, Feng QF. Plant pathogenesis-related proteins and research progress in tobacco [J]. *Biotechnol Bull*, 2012 (5): 89-102]
- 91 周修腾, 王雪, 陈敏, 黄璐琦, 肖文娟, 杨光. 丛枝菌根真菌对丹参木质部结构及防御相关基因的影响[J]. 中国农学通报, 2017, **33** (4): 98-104 [Zhou XT, Wang X, Chen M, Huang LQ, Xiao WJ, Yang G. Effects of arbuscular mycorrhizal fungi on xylem structure and expression of defense related genes in *Salvia miltiorrhiza* [J]. *Chin Agric Sci Bull*, 2017, **33** (4): 98-104]
- 92 谢丽君, 宋圆圆, 曾任森, 王瑞龙, 魏晓晨. 丛枝菌根菌丝桥介导的番茄植株系间抗病信号的传递[J]. 应用生态学报, 2012, **23** (5): 1145-1152 [Xie LJ, Song YY, Ceng RS, Wang RL, Wei XC. Disease resistance signal transfer between roots of different tomato plants through common arbuscular mycorrhizal networks [J]. *Chin J Appl Ecol*, 2012, **23** (5): 1145-1152]
- 93 Mora-Romero GA, Cervantes-Gámez RG, Galindo-Flores H, González-Ortíz MA, Félix-Gastélum R, Maldonado-Mendoza IE, Pérez RS, León-Félix J, Martínez-Valenzuela MC, López-Meyer M. Mycorrhiza-induced protection against pathogens is both genotype-specific and graft-transmissible [J]. *Symbiosis*, 2015, **66** (2): 55-64
- 94 乐仲发. 土著及外源丛枝菌根真菌对三叶赤楠生长效应的研究[D]. 南昌: 江西农业大学, 2015 [Le ZF. Study on the effect of indigenous and exogenous arbuscular mycorrhizal fungi on the growth of *Syzygium grijsii* [D]. Nanchang: Jiangxi Agricultural University, 2015]
- 95 Miller RL, Jackson LE. Survey of vesicular-arbuscular mycorrhizae in lettuce production in relation to management and soil factors. *J Agric Sci*, 1998, **130** (130): 173-182
- 96 田慧, 盖京苹, 李晓林, 张俊伶. 农田土著丛枝菌根真菌群落特征和磷吸收作用研究进展[J]. 土壤通报, 2013, **44** (6): 1512-1519 [Tian H, Gai JP, Li XL, Zhang JL. Community composition and phosphorus uptake by indigenous arbuscular mycorrhizal fungi in agroecosystems [J]. *Chin J Soil Sci*, 2013, **44** (6): 1512-1519]
- 97 王倡宪, 郝志鹏. 丛枝菌根真菌对黄瓜枯萎病的影响[J]. 菌物学报, 2008, **27** (3): 395-404 [Wang CX, Hao ZP. Effects of arbuscular mycorrhizal fungi on *Fusarium* wilt of cucumber seedlings [J]. *Mycosistema*, 2008, **27** (3): 395-404]
- 98 Ren LX, Zhang N, Wu P, Huo H, Xu G. Arbuscular mycorrhizal colonization alleviates *Fusarium* wilt in watermelon and modulates the composition of root exudates [J]. *Plant Growth Regul*, 2015, **77** (1): 77-85
- 99 江龙, 王智明, 张长华, 黄建国, 袁玲. 菌根烟苗的抗青枯病效应研究[J]. 中国烟草学报, 2009, **15** (6): 49-52 [Jiang L, Wang ZM, Zhang CH, Huang JG, Yuan L. Research on wilt disease resistance of AM mycorrhizal tobacco seedlings [J]. *Acta Tabac Sin*, 2009, **15** (6): 49-52]
- 100 Ren LX, Lou YS, Inubushi, K, Sakamoto K, Amemiya Y. Effects of arbuscular mycorrhizal colonization on microbial community in rhizosphere soil and *Fusarium* wilt disease in tomato [J]. *Commun Soil Sci Plant Anal*, 2010, **41** (11): 1399-1410
- 101 张焕仕, 钦佩, 潘少明, 贺学礼. 非灭菌土接种AM真菌对油蒿抗旱性的影响[J]. 北方园艺, 2012 (2): 1-5 [Zhang HS, Qing P, Pan SM, He XL. Effects of AM fungi on drought tolerance of *Artemisia annua* in unsterilized soil [J]. *Northern Horticult*, 2012 (2): 1-5]
- 102 Jakobsen I, Gazey C, Abbott LK. Phosphate transport by communities of arbuscular mycorrhizal fungi in intact soil cores [J]. *New Phytol*, 2001, **149** (1): 95-103
- 103 Daft MJ. The influence of mixed inocula on endomycorrhizal development [J]. *Plant Soil*, 1983, **71** (1-3): 331-337
- 104 Daft MJ, Hogarth BG. Competitive interactions amongst four species of *Glomus* on maize and onion [J]. *Trans Brit Mycol Soc*, 1983, **80** (2): 339-345
- 105 Hu JL, Lin XG, Wang JH, Shen WS, Wu S, Peng SP, Mao TT. Arbuscular mycorrhizal fungal inoculation enhances suppression of cucumber *Fusarium* wilt in greenhouse soils [J]. *Pedosphere*, 2010, **20** (5): 586-593
- 106 龙良鲲, 黎志坤, 姚青, 王燕, 朱红惠. 番茄菌根化育苗及对青枯病的防治试验[J]. 中国蔬菜, 2009, **1** (4): 52-55 [Long LK, Li ZK, Yao Q, Wang Y, Zhu HH. Tomato seedling culture by inoculating arbuscular mycorrhizal fungi and *Ralstonia solanacearum* control [J]. *China Veg*, 2009, **1** (4): 52-55]
- 107 Ashok S, Keerti D, Deepak V, Jha A. Interactions between arbuscular mycorrhizae and *Fusarium oxysporum* f. sp. *ciceris*: effects on fungal development, seedling growth and wilt disease suppression in *Cicer arietinum* L [J]. *Arch Phytopath Plant Prot*, 2015, **48** (3): 240-252
- 108 Barea JM, Azcón-Aguilar C. Microbial co-operation in the rhizosphere [J]. *J Exp Bot*, 2005, **56** (417): 1761-1778
- 109 Behn O. Influence of *Pseudomonas fluorescens*, and arbuscular mycorrhiza on the growth, yield, quality and resistance of wheat infected with *Gaeumannomyces graminis* [J]. *J Plant Dis Prot*, 2008, **115** (1): 4-8
- 110 Akkopru A, Demir S. Biological control of *Fusarium* wilt in tomato

- caused by *Fusarium oxysporum* f. sp. *lycopersici* by AMF *Glomus intraradices* and some rhizobacteria [J]. *J Phytopathol*, 2005, **153** (9): 544-550
- 111 Berta G, Copetta A, Gamalero E, Bona E, Cesaro P. Maize development and grain quality are differentially affected by mycorrhizal fungi and a growth-promoting pseudomonad in the field [J]. *Mycorrhiza*, 2014, **24** (3): 161-170
- 112 Larsen J, Cornejo P, Barea JM. Interactions between the arbuscular mycorrhizal fungus *Glomus intraradices* and the plant growth promoting rhizobacteria *Paenibacillus polymyxa* and *P. macerans* in the mycorrhizosphere of *Cucumis sativus* [J]. *Soil Biol Biochem*, 2009, **41** (2): 286-292
- 113 刘东岳, 李敏, 孙文献, 刘润进. 抗黄瓜枯萎病丛枝菌根真菌与根围促生细菌组合菌剂的筛选[J]. 植物病理学报, 2016, **46** (6): 821-832 [Liu DY, Li M, Sui WX, Liu RJ. Selection of combinations of arbuscular mycorrhizal fungi and plant growth promoting rhizobacteria against cucumber *Fusarium* wilt disease [J]. *Acta Phytopathol Sin*, 2016, **46** (6): 821-832]
- 114 Vestberg M, Kukkonen S, Saari K, Parikka P, Huttunen J. Microbial inoculation for improving the growth and health of micropropagated strawberry [J]. *Appl Soil Ecol*, 2004, **27** (3): 243-258
- 115 Liu R, Dai M, Wu X, Li M, Liu X. Suppression of the root-knot nematode [*Meloidogyne incognita* (Kofoid & White) Chitwood] on tomato by dual inoculation with arbuscular mycorrhizal fungi and plant growth-promoting rhizobacteria [J]. *Mycorrhiza*, 2012, **22** (4): 289-296
- 116 Sharma IP, Sharma AK. Physiological and biochemical changes in tomato cultivar PT-3 with dual inoculation of mycorrhiza and PGPR against root-knot nematode. *Symbiosis*, 2017, **71** (3): 175-183
- 117 Armada E, Probanza A, Roldán A. Native plant growth promoting bacteria *Bacillus thuringiensis* and mixed or individual mycorrhizal species improved drought tolerance and oxidative metabolism in *lavandula dentata* plants [J]. *J Plant Physiol*, 2016, **192**: 1-12
- 118 Velivelli SLS, Lojan P, Cranenbrouck S. The induction of ethylene response factor 3 (ERF3) in potato as a result of coinoculation with *Pseudomonas* sp. R41805 and *Rhizophagus irregularis* MUCL 41833-a possible role in plant defense [J]. *Plant Sign Behav*, 2015, **10** (2): 976-988
- 119 Alizadeh H, Behboudi K, Ahmadzadeh M, Javan-Nikkahet M. Induced systemic resistance in cucumber and *Arabidopsis thaliana*, by the combination of *Trichoderma harzianum* Tr6 and *Pseudomonas* sp. Ps14 [J]. *BioControl*, 2013, **65** (1): 14-23
- 120 刘润进. 菌根真菌是唱响生物共生交响曲的主角[J]. 菌物学报, 2017, **36** (7): 791-799 [Liu RJ. Mycorrhizal fungi are the main factor of singing biological symbiosis symphony [J]. *Mycosistema*, 2017, **36** (7): 791-799]
- 121 Smith SE, Read DJ. Mycorrhizal Symbiosis [M]. 3rd ed. London: Academic Press, 2008
- 122 Eom AH, Hartnett DC, Wilson GWT. Host plant species effects on arbuscular mycorrhizal fungal communities in tallgrass prairie [J]. *Oecologia*, 2000, **122** (3): 435-444
- 123 Cui X, Hu J, Wang J, Yang J, Lin X. Reclamation negatively influences arbuscular mycorrhizal fungal community structure and diversity in coastal saline-alkaline land in eastern China as revealed by Illumina sequencing [J]. *Appl Soil Ecol*, 2016, **98**: 140-149
- 124 Hu JL, Yang AN, Wang JH, Zhu AN, Dai J, Wong MH, Lin XG. Arbuscular mycorrhizal fungal species composition, propagule density, and soil alkaline phosphatase activity in response to continuous and alternate no-tillage in northern China [J]. *Catena*, 2015, **133**: 215-220
- 125 Dai J, Hu JL, Zhu AN, Bai JF. No tillage enhances arbuscular mycorrhizal fungal population, glomalin-related soil protein content, and organic carbon accumulation in soil macroaggregates [J]. *J Soil Sediment*, 2015, **15** (5): 1055-1062
- 126 Wu JH, Shen WS, Lin LM, Greenes RA, Bates DW. Testing the technology acceptance model for evaluating healthcare professionals' intention to use an adverse event reporting system [J]. *Int J Qual Health Care*, 2008, **20** (2): 123-129
- 127 Bainard LD, Klironomos JN, Gordon AM. Arbuscular mycorrhizal fungi in tree-based intercropping systems: a review of their abundance and diversity [J]. *Pedobiologia*, 2011, **54** (2): 57-61
- 128 马琨, 杨桂丽, 马玲, 汪春明, 魏常慧, 代晓华. 间作栽培对连作马铃薯根际土壤微生物群落的影响[J]. 生态学报, 2016, **36** (10): 2987-2995 [Ma K, Yang GL, Ma L, Wang CM, Wei CH, Dai XH. Effects of intercropping on soil microbial communities after long-term potato monoculture [J]. *Acta Ecol Sin*, 2016, **36** (10): 2987-2995]
- 129 Bainard LD, Klironomos JN, Gordon AM. Arbuscular mycorrhizal fungi in tree-based intercropping systems: a review of their abundance and diversity [J]. *Pedobiologia*, 2011, **54** (2): 57-61
- 130 魏常慧. 马铃薯间、套作栽培对连作土壤与作物的影响[D]. 银川: 宁夏大学, 2016 [Wei CH. Effects of intercropping and intercropping on soil and crops in continuous cropping [D]. Yinchuan: Ningxia University, 2016]
- 131 刘俭, 曲继松, 张丽娟, 冯海萍, 杨冬艳. 宁夏旱作区域露地西瓜套作栽培适应性研究[J]. 长江蔬菜, 2012 (20): 49-51 [Liu J, Qu JS, Zhang LJ, Feng HP, Yang DY. Adaptability of watermelon intercropping cultivation in open fields in arid area of Ningxia [J]. *J Changjiang Veg*, 2012 (20): 49-51]
- 132 董鲜, 郑青松, 王敏, 周金燕, 沈其荣. 镐态氮和硝态氮对香蕉枯萎病发生的比较研究[J]. 植物病理学报, 2015, **45** (1): 73-79 [Dong X, Zheng QS, Wang M, Zhou JY, Shen QR. Comparative study of ammonium and nitrate on banana wilt disease development [J]. *Acta Phytopathol Sin*, 2015, **45** (1): 73-79]
- 133 Mäder P, Edelenhofer S, Boller T. Arbuscular mycorrhizae in a long-term field trial comparing low-input (organic, biological) and high-input (conventional) farming systems in a crop rotation [J]. *Biol Fertil Soils*, 2000, **31** (2): 150-156
- 134 Thomson BD, Robson AD, Abbott LK. Soil mediated effects of phosphorus supply on the formation of mycorrhizas by *Scutellispora calospora* (Nicol. & Gerd.) Walker & Sanders on subterranean clover [J]. *New Phytol*, 2010, **118** (3): 463-469
- 135 Kahiluoto H, Ketoja E, Vestberg M, Saarela I. Promotion of AM utilization through reduced P fertilization 2. Field studies [J]. *Plant Soil*, 2001, **231** (1): 65-79
- 136 Martensson AM, Carlgren K. Impact of phosphorus fertilization on VAM diaspores in two Swedish long-term field experiment [J]. *Agric Ecosyst Environ*, 1994, **47** (4): 327-334
- 137 郭涛, 申鸿, 彭思利, 黄建国. 氮、磷供给水平对丛枝菌根真菌生长

- 发育的影响[J]. 植物营养与肥料学报, 2009, **15** (3): 690-695 [Guo T, Shen H, Peng SL, Huang JG. Influence of nitrogen and phosphorus supplying levels on development of arbuscular mycorrhizal fungi [J]. *J Plant Nutr Fertil*, 2009, **15** (3): 690-695]
- 138 刘佳. 长期施肥对农田土壤丛枝菌根真菌生物多样性及群落结构的影响[D]. 兰州: 兰州大学, 2009 [Liu J. Effects of long term fertilization on the diversity and community structure of arbuscular mycorrhizal fungi in Northwest China [D]. Lanzhou: Lanzhou University, 2009]
- 139 林先贵, 胡君利, 戴珏, 王发园, 冯有智. 丛枝菌根真菌群落结构与多样性研究方法概述及实例比较[J]. 应用与环境生物学报, 2017, **23** (2): 343-350 [Lin XG, Hu JL, Dai J, Wang FY, Feng YZ. Overview and comparison of research methods for determining the community structure and diversity of arbuscular mycorrhizal fungi [J]. *Chin J Appl Environ Biol*, 2017, **23** (2): 343-350]
- 140 Johnson NC. Can fertilization of soil select less mutualistic mycorrhizae [J] *Ecol Applic*, 1993, **3** (4): 749-757
- 141 Wang MY, Hu LB, Wang WH, Liu ST, Li M, Liu RJ. Influence of long term fixed fertilization on diversity of arbuscular mycorrhizal fung [J]. *Pedosphere*, 2009, **19** (5): 663-672
- 142 Beuregard MS, Hamel C, Atul-Nayyar, St-Arnaud M. Long term phosphorus fertilization impacts soil fungaland bacterial diversity but not AM fungal community in alfalfa [J]. *Microb Ecol*, 2010, **59** (2): 379-389
- 143 keitaro T, Ryouta H, Tadao W. Inoculation of arbuscular fungi can substantially reduce phosphate fertilizer application to *Allium fistulosum* L. and achieve marketable yield under field condition [J]. *Biol Fertil Soils*, 2012, **48** (7): 839-843
- 144 赵慧敏. 丛枝菌根生理生态学研究进展[J]. 安徽农业科学, 2009, **37** (4): 1460-1462 [Zhao HM. Research progress on eco-physiological responses of arbuscular mycorrhizal [J]. *J Anhui Agric Sci*, 2009, **37** (4): 1460-1462]
- 145 李晓林, 冯固. 丛枝菌根生态生理. 北京: 华文出版社, 2001 [Li XL, Feng G. Eco-phisiology of Arbuscular Mycorrhiza. Beijing: Sino-Culture Press, 2001]
- 146 赵方贵, 刘广富, 管志坤, 朱柱峰, 刘广玉. 土壤根际环境对烟草AM真菌定殖率和孢子密度的影响[J]. 安徽农业科学, 2012 (7): 4220-4221 [Zhao FG, Liu GF, Guan ZK, Zhu ZF, Liu GY. Influence of soil chemical and physical properties on AM fungi infections and spore density of tobacco in Shandong [J]. *J Anhui Agric Sci*, 2012 (7): 4220-4221]
- 147 袁飞, 彭宇, 张春兰. 有机物料减轻设施连作黄瓜苗期病害的微生物效应[J]. 应用生态学报, 2004, **15** (5): 867-870 [Yuan F, Peng Y, Zhang CL. Microbial effects of organic material mitigation facilities continuous cropping on cucumber seedling disease [J]. *Chin J Appl Ecol*, 2004, **15** (5): 867-870]
- 148 李军营, 邓小鹏, 杨坤. 施用有机肥对植烟土壤理化性质的影响[J]. 中国土壤与肥料, 2012 (3): 12-16 [Li JY, Deng XP, Yang K. Effects of application of organic fertilizer on the physical and chemical properties of tobacco planting soil [J]. *Soil Fertil Sci China*, 2012 (3): 12-16]
- 149 李灵芝, 行园园, 盖京苹, 王艳芳, 李海平. 不同氮肥及有机肥投入对设施番茄菜地AM真菌侵染势和空间分布的影响[J]. 中国蔬菜, 2015, **1** (4): 49-53 [Li LZ, Hang YY, Gai JP, Wang YF, Li HP. Effects of different nitrogen and organic manure application on infection potential and spatial distribution of arbuscular mycorrhizal fungi in greenhouse tamato cropping system [J]. *China Veg*, 2015, **1** (4): 49-53]
- 150 Wang MY, Hu LB, Wang WH. Influence of long-term fixed fertilization on diversity of arbuscular mycorrhizalfungi [J]. *Pedosphere*, 2009, **19** (5): 663-672
- 151 Hodge A, Campbell CD, Fitter AH. An arbuscular mycorrhizal fungus accelerates decomposition and acquires nitrogen directly from organic material [J]. *Nature*, 2001, **413** (6853): 297-299
- 152 Verma RK, Arya ID. Effect of arbuscular mycorrhizal fungal isolates and organic manure on growth and mycorrhization of micropropagated *Dendrocalamus asper*, plantlets and on spore production in their rhizosphere [J]. *Mycorrhiza*, 1998, **8** (2): 113-116
- 153 He XL, Steinberger Y, Herman P. Dynamics on arbuscular mycorrhizal fungi under the canopy of desert shrubs [J]. *J NW A&F Univ Nat Sci*, 2001, **29** (4): 24-28
- 154 Dai J, Hu JL, Lin XG, Yang AN. Arbuscular mycorrhizal fungal diversity, external mycelium length, and glomalin-related soil protein content in response to long-term fertilizer management [J]. *J Soil Sediment*, 2013, **13** (1): 1-11
- 155 Hu J, Lin X, Wang J, Chu H, Yin R, Zhang J. Population size and specific potential of P-mineralizing and -solubilizing bacteria under long-term P-deficiency fertilization in a sandy loam soil [J]. *Pedobiologia*, 2009, **53** (1): 49-58
- 156 张贵云. 不同农业措施对丛枝菌根真菌群落结构和侵染效应的影响[D]. 南京: 南京农业大学, 2013 [Zhang GY. Effects of different agricultural measures on the community structure and infection effect of arbuscular mycorrhizal fungi [D]. Nanjing: Nanjing Agricultural University, 2013]
- 157 Khaosaad T, García-Garrido JM, Steinkellner S, Vierheilig H. Take-all disease is systemically reduced in roots of mycorrhizal barley plants [J]. *Soil Biol Biochem*, 2007, **39** (3): 727-734