

肿瘤治疗性mRNA疫苗的研发进展

邓可欣^{1,2†}, 李晓彬^{1,2†}, 刘婉婉^{1,2}, 刘君禹^{1,2}, 王润铭^{1,2,3*}, 尹延东⁴, 张灿阳^{1,2,3,4*}

1. 清华大学深圳国际研究生院生物医药与健康工程研究院, 深圳 518055

2. 清华大学深圳国际研究生院活性蛋白多肽绿色生物制造广东普通高校重点实验室, 深圳 518055

3. 清华大学工业生物催化教育部重点实验室, 北京 100084

4. 深圳湾实验室, 深圳 518132

† 同等贡献

* 联系人, E-mail: rungmingwang@sz.tsinghua.edu.cn; zhang.cy@sz.tsinghua.edu.cn

2024-08-18 收稿, 2024-10-12 修回, 2024-12-02 接受, 2024-12-03 网络版发表

广东省珠江人才计划(2021QN02Y225)、清华大学深圳国际研究生院海外科研合作基金(HW2023009)和化工系-iBHE专项合作联合基金资助

摘要 近年来, 肿瘤疫苗作为肿瘤免疫疗法的重要组成部分, 已经取得了显著的进展。肿瘤治疗性mRNA疫苗通过递送肿瘤相关抗原或肿瘤特异性抗原, 激发机体产生特异性免疫反应, 以识别并杀伤肿瘤细胞。相较于其他类型的肿瘤疫苗, mRNA疫苗因其独特的优势, 在临床试验中展现出良好的治疗效果和巨大的应用潜力。mRNA疫苗的优势在于其快速的开发周期、高度的特异性, 以及能够激发强烈的免疫反应。它们不整合入宿主基因组, 降低了安全性风险, 同时可以快速应对病原体的变异。此外, mRNA疫苗的稳定性可以通过特定的修饰来提高, 增强其在体内的持久性和翻译效率, 从而增强疫苗的效果。目前, 多种个体化mRNA肿瘤疫苗在临床试验中表现出较好的安全性和免疫原性, 显示了其作为肿瘤治疗工具的潜力。本文总结了肿瘤治疗性mRNA疫苗的构成、优势、稳定性提高方法、作用机制、给药途径、递送系统、局限性和挑战等, 旨在促进肿瘤治疗性mRNA疫苗的发展和应用。

关键词 mRNA疫苗, 递送系统, 给药途径, 作用机制, 肿瘤免疫

肿瘤免疫治疗是癌症治疗的新兴领域, 肿瘤治疗性疫苗作为肿瘤免疫治疗的重要组成部分, 是未来癌症治疗领域最具前景的候选药物^[1~3]。肿瘤治疗性疫苗通过刺激机体针对特异性肿瘤抗原产生适应性免疫, 放大并维持特异性T细胞响应来杀死肿瘤细胞, 从而达到控制肿瘤生长、延长患者生存期的目的^[4,5](图1), 同时不同的给药方式会影响治疗效果, 结合不同的递送体系可以实现特异性治疗, 提高治疗效果。

肿瘤治疗性疫苗包括重组蛋白/多肽疫苗、细胞疫苗、DNA/mRNA疫苗、细菌和病毒载体疫苗^[6,7]。其中, mRNA疫苗因其独特的优势而备受关注。mRNA疫苗的核心组成部分是编码肿瘤抗原的mRNA序列, 这些序列通过特定的递送载体进入宿主细胞, 并指导细

胞产生相应的肿瘤抗原蛋白, 从而激活免疫系统。一个典型的mRNA疫苗分子包括5'帽子结构, 5'非翻译区(5'UTR)、开放阅读框(ORF)、3'非翻译区(3'UTR)、以及3'Poly(A)尾部。这些结构元件共同确保mRNA的稳定性、翻译效率以及免疫原性^[8,9]。基于mRNA的结构, mRNA肿瘤治疗性疫苗主要分为非复制型mRNA疫苗、自扩增mRNA疫苗和反扩增mRNA疫苗。mRNA疫苗用于肿瘤治疗具有许多优势, 首先, 机体对mRNA疫苗的耐受性良好, mRNA肿瘤治疗性疫苗产生的药物不良反应通常是可控且短暂的^[10~13]; 其次, 与DNA疫苗相比, mRNA疫苗没有插入宿主基因组引起突变的风险^[9,14]; 与细菌或病毒载体疫苗相比, mRNA治疗性疫苗不使用致病性细菌或病毒制剂, 具有非传染性^[15];

引用格式: 邓可欣, 李晓彬, 刘婉婉, 等. 肿瘤治疗性mRNA疫苗的研发进展. 科学通报, 2025, 70: 432–442

Deng K, Li X, Liu W, et al. Advances in the development of therapeutic mRNA vaccines for cancer therapy (in Chinese). Chin Sci Bull, 2025, 70: 432–442,
 doi: [10.1360/TB-2024-0878](https://doi.org/10.1360/TB-2024-0878)

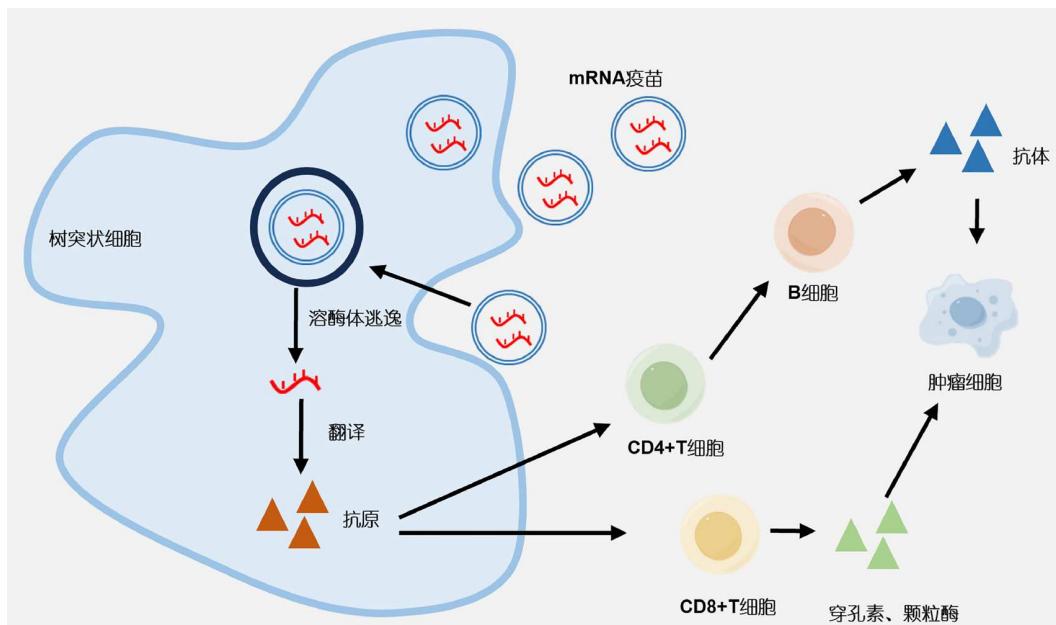


图 1 (网络版彩色)肿瘤治疗性mRNA疫苗的作用机制

Figure 1 (Color online) Mechanism of therapeutic mRNA vaccines for cancer treatment

另外,相较于直接注射抗原, mRNA疫苗能够使人体持续高效地形成自发性的体液和细胞免疫;最后, mRNA可通过体外转录技术合成,生产过程简单快速,生产成本较低^[9,16,17]。

保证mRNA的完整性和稳定性是合成有效的肿瘤mRNA治疗性疫苗的关键^[18,19]。从mRNA的结构上讲,mRNA的5'帽子结构是真核生物信使核糖核酸的特征,有助于核糖体识别和翻译^[20~23]。体外转录技术获得的mRNA具有无帽或异常的帽子结构,被模式识别受体(pattern recognition receptors, PRRs)识别,从而刺激I型干扰素产生,干扰mRNA翻译,因此体外转录技术获得的mRNA应进行适当封端,以防被识别为外来抗原^[20,24]。mRNA的Poly(A)尾部也影响mRNA的翻译效率和稳定性, Poly(A)尾部能够降低RNA核酸外切酶的活性,从而有助于提高mRNA的翻译效率和稳定性^[25~27]。Poly(A)尾部的长度应控制在100~300个核苷酸,长度对调控mRNA的翻译和稳定性起着重要作用,Poly(A)尾部的长度与翻译效率成正比^[28]。体外转录技术合成的mRNA可以通过DNA模板合成或使用Poly(A)聚合酶在转录后通过酶促反应添加Poly(A)尾^[6,24]。UTR元件是mRNA结构设计的关键部分,5'UTR对靶蛋白的表达至关重要,3'UTR影响mRNA的半衰期,因此,修饰5'和3'UTR能够有效提高体外转录mRNA翻译效

率和半衰期^[15,29]。此外,通过假尿苷、1-甲基假尿苷和5-甲基胞嘧啶等对mRNA进行适当修饰能够有效提高mRNA的稳定性和翻译效率^[30]。体外转录技术合成的mRNA具有许多siRNA、dsRNA等副产物,这些副产物能够激活体内PRRs,阻断mRNA翻译,并诱导先天性免疫反应,因此,需要对体外转录技术合成的mRNA进行纯化以减少免疫刺激^[31,32]。

1 肿瘤治疗性mRNA的作用机制

当mRNA疫苗在给药部位附近被抗原呈递细胞(antigen presenting cell, APC)内吞,内体化的mRNA转运到细胞质核糖体中进行翻译,翻译产生的相关抗原蛋白随后进行如泛素化等翻译后修饰。蛋白酶体复合物将抗原蛋白降解成为小片段,通过主要组织相容性复合物(major histocompatibility complex, MHC) I类分子进行两种类型的抗原呈递:(1)部分小片段肽通过抗原处理相关转运蛋白转运到糙面内质网上,由MHC I类分子呈递到细胞表面,活化的CD8⁺ T细胞识别到相关抗原后,分泌穿孔素、颗粒酶等分子诱导肿瘤细胞凋亡^[33]。(2)分泌的抗原可被细胞摄取,随后被降解,并由MHC II类分子呈递给辅助性T细胞。辅助性T细胞可以激活B细胞产生中和抗体,并通过炎症因子激活吞噬细胞,最终清除肿瘤细胞^[34]。

在肿瘤抗原的选择上,肿瘤治疗性mRNA疫苗的抗原通常包括肿瘤相关抗原(tumor-associated antigens, TAAs)(表1)和肿瘤特异性抗原(tumor-specific antigens, TSAs)(表2)^[32,35]。TAAs通常在肿瘤细胞中过表达,肿瘤特异性和免疫原性较弱。肿瘤相关自身抗原是目前最常采用的肿瘤抗原,但哺乳动物对单一的TAAs具有较高的免疫耐受性,因此目前通常采用多种TAAs联合应用的方法^[6,36]。TSAs是来源于肿瘤细胞突变的肿瘤新抗原,正常组织不表达,具有较高的肿瘤特异性和免疫原性,但机体耐受性较弱^[37]。通过癌症患者肿瘤细胞内独特的突变特征可设计出个性化定制的肿瘤疫苗,目前临幊上已经开发了多种编码TSAs的个性化疫苗,但由于TSAs的特殊性,个性化肿瘤疫苗的成本较大,这极大限制了基于TSAs肿瘤疫苗的开发和应用^[38,39]。

2 肿瘤治疗性mRNA疫苗的给药途径

不同的给药途径会影响mRNA肿瘤治疗性疫苗的免疫效力及刺激响应区域,目前mRNA肿瘤治疗性疫苗的给药途径主要包括静脉注射、肌内注射、皮下注

射、经鼻给药、皮下注射等^[9](表3)。

2.1 静脉注射

静脉注射(intravenous injection)是目前治疗性mRNA肿瘤疫苗临床试验中较常采用的给药方式。静脉给药能使疫苗达到多个淋巴器官,高效诱导对肿瘤细胞的免疫反应^[40-42]。通过静脉注射mRNA疫苗BNT111能有效诱导黑色素瘤患者体内CD4⁺ T细胞和CD8⁺ T细胞免疫应答^[43],但可能会导致全身性的副作用,如脾损伤和淋巴细胞耗竭。

2.2 肌内注射

肌内注射(intramuscular injection)是将疫苗直接注射到肌肉组织中,操作方便、副作用小且耐受性好,是一种广泛使用且可行的注射方式,但是剂量有限,容易引起局部疼痛、红肿。目前批准上市的SARS-CoV-2 mRNA疫苗采用的是肌内注射^[44-46]。Chen等人^[47]报道了通过肌内注射基于iso-A11B5C1脂质的疫苗能够显著减缓B16F10黑色素瘤的生长且肿瘤组织表现出显著

表1 TAA联用疫苗

Table 1 TAA combination vaccines

疫苗名称	针对肿瘤类型	抗原	研发公司
CV9201	非小细胞肺癌	NY-ESO-1、MAGE-C1/C2 5T4	德国CureVac
CV9202	非小细胞肺癌	MAGE-A3、MAGE-C1、MAGE-C2、5T4、Survivin和MUC-1	德国CureVac
BNT111	黑色素瘤	NY-ESO-1、MAGE-A3、酪氨酸酶和TPTE	德国BioNtech

表2 代表性TSA疫苗

Table 2 TSA-based vaccines

疫苗名称	针对肿瘤类型	抗原	研发公司
mRNA-5671	广谱实体瘤	G12D、G12V、G13D、G12C	美国Moderna
mRNA-4157	黑色素瘤	34种病患特异性新抗原	美国Moderna
BNT122	胰腺导管腺癌	20种病患特异性MHC I类和II类限制新抗原	德国BioNtech
CV9103	前列腺癌	PSA、PSCA、PAP、PSMA	德国CureVac

表3 不同mRNA肿瘤治疗性疫苗给药途径对比表

Table 3 Comparison of different mRNA therapeutic vaccine delivery routes for tumor

给药途径	生物利用度	免疫应答强度	安全性	可能的副作用
静脉注射	高	强	较高	静脉炎、全身反应
肌内注射	中	中	高	局部疼痛、红肿
经鼻给药	中	中	高	鼻腔刺激、吸入风险
瘤内注射	高	强	中	注射部位感染、局部反应
皮下注射	中	中	中	局部肿胀、疼痛、过敏反应

的CD8⁺ T细胞浸润。

2.3 经鼻给药

经鼻给药(intranasal administration)属于非侵入性给药, 可以将mRNA疫苗有效地递送到外周淋巴结的抗原呈递细胞, 但受限于鼻腔体积, 给药剂量较小, 容易刺激鼻腔^[48~50]。Mai等人^[51]设计了一种经鼻给药的阳离子脂质体/鱼精蛋白复合物LPC, 该疫苗在体内经鼻给药后显示出强烈的细胞免疫反应并能够明显减缓小鼠肿瘤生长。

2.4 瘤内注射

瘤内注射(intratumoral injection)mRNA疫苗可以快速激活免疫细胞并最大限度地减少药物脱靶, 但瘤内给药有效性受到肿瘤大小和位置的限制, 还容易产生局部强烈的免疫反应^[52~54]。Deng等人^[55]设计并优化了编码OX40L的mRNA疫苗, 该疫苗通过瘤内注射给药后能够显著抑制肿瘤生长, 提高小鼠存活率并增加疫苗给药组小鼠体内的CD4⁺ T细胞和CD8⁺ T细胞水平。

2.5 皮下注射

皮下注射(subcutaneous injection)mRNA疫苗易被局部区域的抗原呈递细胞处理, 同时容易实现大剂量给药, 但皮下给药往往会引起较大的局部注射部位反应^[24,56,57]。

3 肿瘤治疗性mRNA疫苗的递送系统

由于mRNA分子量比较大且带有负电荷, 游离的mRNA无法穿过同样带有负电荷的磷脂双分子层膜, 进入细胞比较困难^[58]。在体内, 游离的mRNA容易被机体先天性免疫系统识别吞噬, 且很难实现溶酶体逃逸, 体内广泛存在的核酸酶容易将mRNA降解, 因此需要合适的递送载体, 能将mRNA完整高效地递送到体内, 并能根据实际需求应用于多种场景^[34,59]。目前, 针对肿瘤治疗性mRNA疫苗已经开发出多种创新型的递送系统, 如基于脂质的递送、基于聚合物的递送、基于树突状细胞的递送等, 能够有效提高mRNA的递送效率和稳定性^[33,34](图2)。

3.1 基于脂质的递送系统

脂质纳米颗粒是目前最成熟的mRNA递送体系, 批准上市的治疗新冠病毒mRNA疫苗使用脂质纳米颗

粒(lipid nanoparticles, LNPs)作为递送载体^[60~62]。一般来说, 脂质纳米颗粒由四种主要组分构成: 阳离子或可电离脂质、辅助性中性脂质、胆固醇和聚乙二醇(polyethylene glycol, PEG)化脂质。其中, 阳离子或可电离脂质是LNPs的主要成分, 通过静电相互作用吸附mRNA, 促进封装, 有利于mRNA疫苗完成溶酶体逃逸, 同时能提高mRNA转染效率^[63]。常用的阳离子脂质如DOTAP、DOTMA等。常见的可电离脂质如SM-102、Dlin-MC3-DMA、ALC-0315等^[34,64]。辅助性中性脂质如DOPE、DSPC等有助于提高脂质纳米颗粒的稳定性, 促进核酸封装^[65,66]。胆固醇嵌入磷脂分子中, 可以调节LNPs磷脂双分子层膜的流动性, 稳定纳米材料, 减少药物渗漏^[67]。聚乙二醇化脂质如ALC-0159、PEG-DMG是LNPs的重要组成部分, PEG在脂质体纳米颗粒表面, 通过大分子空间位阻减少LNPs的聚集, 同时降低脂质体和血浆蛋白的相互吸附, 延长脂质体在体内的循环时间, 另一方面, 可根据需求将脂质体通过PEG进行改造, 达到提高生物相容性和稳定性等目的^[68]。

目前, 许多肿瘤治疗性mRNA疫苗的研究主要采用脂质纳米颗粒作为递送系统。目前还没有上市的治疗肿瘤的mRNA疫苗, 基于脂质体纳米颗粒的肿瘤疫苗mRNA-4157目前进入临床三期研究(NCT03897881), 该研究以完全切除皮肤黑色素瘤且复发风险高的受试者作为对象, 旨在比较mRNA-4157与帕博利珠单抗(pembrolizumab)联合或单药治疗在肿瘤患者术后辅助治疗中对无复发生存率(recurrence free survival, RFS)的影响。mRNA-4157是一款个性化的肿瘤疫苗, 可编码多达34种肿瘤新抗原, 疫苗的前期临床结果表明, 与单药治疗相比, 联合治疗的无复发生存期更长^[69]。以脂质为递送系统的肿瘤治疗性mRNA疫苗在三阴性乳腺癌、黑色素瘤、肺癌等肿瘤的研究也有许多进展, 如Liu等人^[70]基于脂质合成LCP纳米颗粒来递送编码肿瘤相关抗原1型跨膜黏蛋白(mucins 1, MUC1)的mRNA, 该mRNA疫苗能够有效诱导针对三阴性乳腺癌的抗原特异性免疫反应, 并和抗细胞毒性T淋巴细胞相关蛋白4(cytotoxic T-lymphocyte-associated protein 4, CTLA-4)的单克隆抗体联合治疗可显著增强疫苗的免疫反应, 抑制肿瘤生长。Chen等人^[71]设计了一种基于脂质纳米颗粒的淋巴结靶向mRNA疫苗113-O12B, 该疫苗的靶向递送可引发强大的CD8⁺ T细胞响应, 治疗小鼠黑色素瘤效果明显。Ma等人^[72]开发了一种mPLA/mRNA(mLPR)肿瘤疫苗, 鼻腔给药后, mLPR疫苗刺激树突状

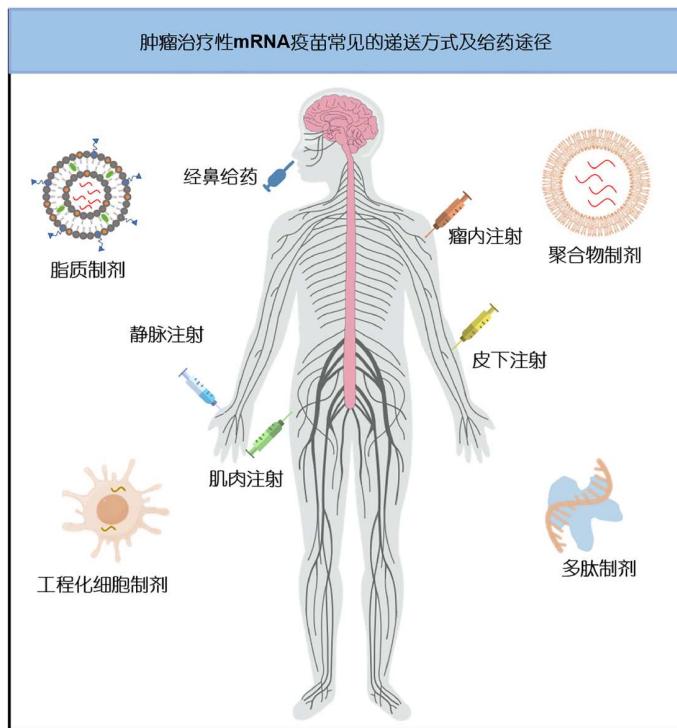


图 2 (网络版彩色)肿瘤治疗性mRNA疫苗常见的递送方式及给药途径

Figure 2 (Color online) Delivery methods and administration routes of therapeutic mRNA vaccines for cancer treatment

细胞成熟，进而通过激活IFN- γ 等细胞因子分泌和自然杀伤细胞介导的抗体依赖性细胞毒性(antibody-dependent cell-mediated cytotoxicity, ADCC)来抑制肿瘤发展并减少骨转移。

3.2 基于聚合物的递送系统

聚合物作为核酸递送平台具有巨大的应用前景，与基于脂质的递送系统相比，聚合物递送载体稳定性及可控性更好，可以延长疫苗体内循环时间、实现组织或细胞靶向^[30,73]。常见的聚合物递送载体包括聚乙烯亚胺(polyethyleneimine, PEI)、聚酯(polyesters)、聚氨基酸(poly-amino acids)和壳聚糖(chitosan)等糖类^[74~76]。其中，聚乙烯亚胺应用较为广泛，PEI具有良好的“质子海绵效应”，可以高效促进溶酶体逃逸，同时，PEI还可以激活树突状细胞、促进细胞因子产生^[77]。Li等人^[78]合成了递送mRNA的材料氟接枝聚乙烯亚胺材料(F-PEI)，F-PEI能够促进核酸递送并激活TLR4介导的信号通路。F-PEI和编码肿瘤相关抗原的mRNA自组装形成纳米疫苗，能刺激树突状细胞成熟进而呈递抗原，引起抗肿瘤免疫反应。Yin等人^[79]研究了一种注射用水

凝胶，由氧化石墨烯和PEI构成，包裹编码卵清蛋白mRNA和佐剂R848，皮下注射至少30天后形成纳米疫苗，该疫苗可靶向淋巴结，刺激CD8⁺ T细胞生成，杀伤肿瘤细胞。

3.3 基于树突状细胞的递送系统

树突状细胞(dendritic cells, DCs)能够通过多种机制摄取和呈递肿瘤相关抗原(TAAs)，从而引发肿瘤特异性免疫反应。此外，树突状细胞还可以在淋巴和非淋巴组织之中迁移、调节细胞因子和炎症趋化因子等，在维持系统和持久的抗肿瘤反应中发挥着重要作用^[80,81]。树突状细胞作为最有效的抗原呈递细胞，一直是肿瘤治疗性疫苗的开发重点，这种疫苗疗法通常通过从外周血中分离出单核细胞或造血干细胞/祖细胞，用重组细胞因子诱导成熟，并负载各种TAAs，最后将成熟的DC疫苗回输到患者体内发挥作用^[82~85]。2010年4月，美国FDA批准治疗转移性去势抵抗性前列腺癌的肿瘤疫苗Sipuleucel-T (provenge; dendreon)上市^[86]。Sipuleucel-T的活性成分包括自体外周血APCs和重组人源PAP-GM-CSF^[87,88]。虽然基于树突状细胞的肿瘤

mRNA治疗性疫苗应用前景良好，但与基于脂质等的mRNA疫苗相比，树突状细胞的来源及体外培养激活等操作既繁琐又费时耗力^[89]。

3.4 其他

阳离子肽鱼精蛋白在mRNA递送领域受到广泛研究，主要通过静电相互作用实现对mRNA的包封，从而有效避免其被核酸酶降解。此外，鱼精蛋白-mRNA复合物还可作为一种佐剂，引发Th-1型免疫反应^[15,90,91]。阳离子细胞穿透肽(cationic cell-penetrating peptides, CPPs)在mRNA疫苗研究中也显示了不错的疗效。Udhayakumar等人^[92]报道了一种递送mRNA的阳离子穿透肽，形成的纳米复合物促进mRNA内体逃逸，进而在DC细胞内表达，且具有pH依赖性的膜破坏特性。

4 进展与展望

近年来，肿瘤治疗性mRNA疫苗在抗原选择、给药途径、递送系统等方面都得到了快速的优化和实质性进展，但仍存在许多问题和挑战^[1,30]。尽管采用TSAs能很大程度地提高mRNA疫苗的免疫原性，但由于肿瘤免疫微环境的复杂性、基因变异的随机性和个体之间的异质性，识别有效的肿瘤特异性突变仍很困难，且大规模生产安全稳定的个性化mRNA疫苗仍有技术壁垒^[4,93-95]。目前肿瘤治疗性mRNA疫苗需要多次给药来加大疫苗效力从而诱导肿瘤免疫反应，这对治疗早期

癌症患者较为有效，但晚期癌症患者体内免疫微环境被高度抑制，单一mRNA疫苗治疗效果可能不尽如人意，目前更加关注肿瘤治疗性mRNA疫苗和传统治疗方法如化疗、放疗和其他免疫疗法的联合治疗(图3)。mRNA-4157和帕博利珠单抗联合治疗与单用帕博利珠单抗治疗相比，联合用药的患者疾病复发风险显著降低^[69,96]。Fotin-Mleczek等人^[97]将mRNA疫苗和放疗结合，表现出强大的抗肿瘤协同作用，联合治疗显著增加了小鼠肿瘤的CD4⁺ T细胞、CD8⁺ T细胞和NKT细胞浸润。Liu等人^[70]构建了编码肿瘤抗原MUC1的mRNA疫苗，并和抗CTLA-4单克隆抗体联用，结果表明，与单药治疗相比，联合疗法可以显著增强抗肿瘤免疫应答。Sahin等人^[43]报道BNT111疫苗Ⅱ期临床实验数据，结果表明，治疗晚期黑色素瘤的肿瘤mRNA疫苗BNT111和抗PD-1抗体西普丽珠单抗联用显示出良好的抗肿瘤反应和安全性。Awad等人^[98]将个性化肿瘤新抗原mRNA疫苗NEO-PV-01与晚期非鳞状非小细胞肺癌(non-small cell lung cancer, NSCLC)一线治疗药物培美曲塞、卡铂和帕博利珠单抗联用，38名治疗患者没有出现与治疗相关的严重不良反应事件，且接种疫苗后观察到抗原特异性的T细胞反应，证实了其在NSCLC中具有较好的安全性和治疗效果。

肿瘤治疗性mRNA疫苗作为一种新兴的治疗手段，近年来在癌症治疗领域取得了显著进展。这些疫苗通过利用mRNA分子编码肿瘤特异性抗原，激发机体的

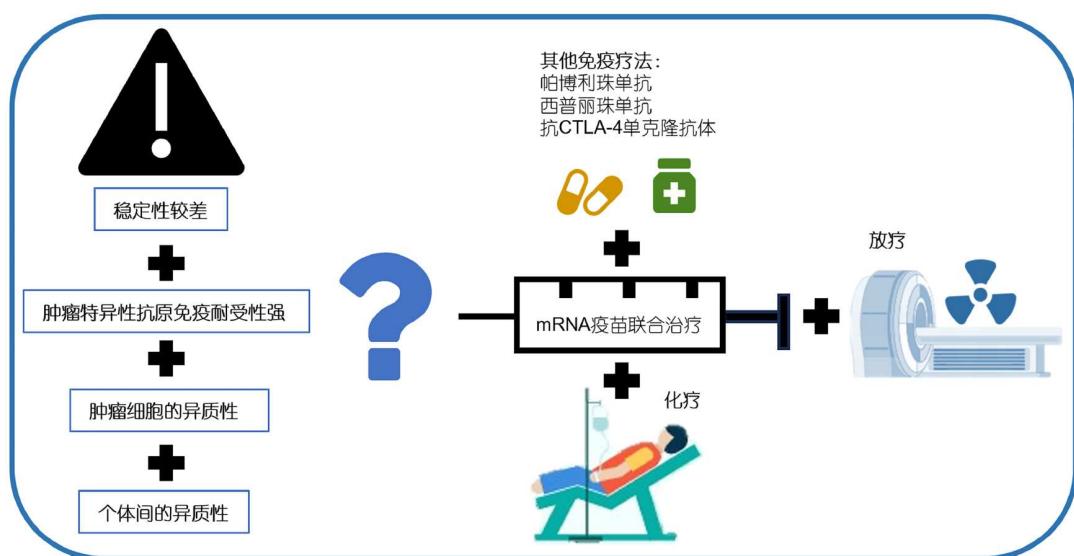


图 3 (网络版彩色)肿瘤治疗性mRNA疫苗的挑战及联合治疗策略

Figure 3 (Color online) Challenges and combination therapy strategies of therapeutic mRNA vaccines for cancer treatment

免疫系统识别并攻击肿瘤细胞。尽管许多研究仍处于临床试验阶段，但初步结果表明，mRNA疫苗在提高患者生存率和生活质量方面具有巨大潜力。mRNA疫苗的优势在于其设计和生产的灵活性，能够快速响应肿瘤抗原的变化，以及其非整合性，降低了插入突变的风险。此外，mRNA疫苗能够诱导强烈的免疫反应，包括激活T细胞和产生特异性抗体。这些特性使得mRNA疫苗成为癌症治疗中一个有前景的研究方向。然而，mRNA疫苗的研究和应用也面临一些挑战，包括如何提高mRNA的稳定性和递送效率，以及如何克服机体

可能产生的免疫耐受。为了解决这些问题，研究人员正在探索多种策略，如优化mRNA序列、开发新型递送系统，以及结合其他治疗方法（如免疫检查点抑制剂）来增强疫苗的效果。随着技术的不断进步和临床试验的深入，mRNA疫苗有望在癌症治疗中发挥更加重要的作用。预计mRNA疫苗将不仅限于治疗性应用，还可能扩展到预防性疫苗的开发，为癌症患者提供更多的治疗选择和更好的治疗效果。此外，mRNA疫苗的研究也可能推动其他领域，如传染性疾病和遗传性疾病的治疗，开启医学治疗的新篇章。

参考文献

- 1 Lorentzen C L, Haanen J B, Met Ö, et al. Clinical advances and ongoing trials of mRNA vaccines for cancer treatment. *Lancet Oncol*, 2022, 23: e450–e458
- 2 Sahin U, Türeci Ö. Personalized vaccines for cancer immunotherapy. *Science*, 2018, 359: 1355–1360
- 3 Liu J, Fu M, Wang M, et al. Cancer vaccines as promising immuno-therapeutics: Platforms and current progress. *J Hematol Oncol*, 2022, 15: 28
- 4 Yuan Y, Gao F, Chang Y, et al. Advances of mRNA vaccine in tumor: A maze of opportunities and challenges. *Biomark Res*, 2023, 11: 6
- 5 Tran T, Blanc C, Granier C, et al. Therapeutic cancer vaccine: Building the future from lessons of the past. *Semin Immunopathol*, 2019, 41: 69–85
- 6 He Q, Gao H, Tan D, et al. mRNA cancer vaccines: Advances, trends and challenges. *Acta Pharm Sin B*, 2022, 12: 2969–2989
- 7 Zhang M M, Li G, Hou T L, et al. Advancements in nanotechnology-enabled mRNA delivery systems (in Chinese). *Chin Sci Bull*, 2024, 69: 4858–4873 [张苗苗, 李港, 侯泰霖, 等. 基于纳米技术的mRNA递送系统的研究进展. 科学通报, 2024, 69: 4858–4873]
- 8 Jackson N A C, Kester K E, Casimiro D, et al. The promise of mRNA vaccines: A biotech and industrial perspective. *npj Vaccines*, 2020, 5: 11
- 9 Hu Y X, Pu C T, Liu B X, et al. The rational design of mRNA vaccine: From empirical method to artificial intelligence-based design (in Chinese). *Chin Sci Bull*, 2024, 69: 4805–4812 [胡宇轩, 濮澄韬, 刘博翔, 等. mRNA疫苗理性设计: 从经验到人工智能设计. 科学通报, 2024, 33: 4805–4812]
- 10 Karikó K, Muramatsu H, Welsh F A, et al. Incorporation of pseudouridine into mRNA yields superior nonimmunogenic vector with increased translational capacity and biological stability. *Mol Ther*, 2008, 16: 1833–1840
- 11 Bird L. mRNA vaccine for treating pancreatic cancer. *Nat Rev Immunol*, 2023, 23: 413
- 12 Zhang A, Ji Q, Sheng X, et al. mRNA vaccine in gastrointestinal tumors: Immunomodulatory effects and immunotherapy. *Biomed Pharmacother*, 2023, 166: 115361
- 13 Wei L, Xue Y C. Breakthroughs in mRNA vaccines and innovations in drug development (in Chinese). *Chin Sci Bull*, 2023, 68: 4948–4953 [魏绿, 薛愿超. mRNA疫苗的突破与药物研发革新. 科学通报, 2023, 36: 4948–4953]
- 14 Thess A, Grund S, Mui B L, et al. Sequence-engineered mRNA without chemical nucleoside modifications enables an effective protein therapy in large animals. *Mol Ther*, 2015, 23: 1456–1464
- 15 Miao L, Zhang Y, Huang L. mRNA vaccine for cancer immunotherapy. *Mol Cancer*, 2021, 20: 41
- 16 Bongianino R, Denegri M, Mazzanti A, et al. Allele-specific silencing of mutant mRNA rescues ultrastructural and arrhythmic phenotype in mice carriers of the R4496C mutation in the ryanodine receptor gene (*RYR2*). *Circ Res*, 2017, 121: 525–536
- 17 Ulmer J B, Mason P W, Geall A, et al. RNA-based vaccines. *Vaccine*, 2012, 30: 4414–4418
- 18 Gote V, Bolla P K, Kommineni N, et al. A comprehensive review of mRNA vaccines. *Int J Mol Sci*, 2023, 24: 2700
- 19 Kon E, Elia U, Peer D. Principles for designing an optimal mRNA lipid nanoparticle vaccine. *Curr Opin Biotechnol*, 2022, 73: 329–336
- 20 Kim S C, Sekhon S S, Shin W R, et al. Modifications of mRNA vaccine structural elements for improving mRNA stability and translation efficiency. *Mol Cell Toxicol*, 2022, 18: 1–8
- 21 Henderson J M, Ujita A, Hill E, et al. Cap 1 messenger RNA synthesis with co-transcriptional cleancap® analog by *in vitro* transcription. *Curr Protocols*, 2021, 1: e39
- 22 Deviatkin A A, Simonov R A, Trutneva K A, et al. Cap-independent circular mRNA translation efficiency. *Vaccines*, 2023, 11: 238
- 23 Liu C, Peng J Y, Zhang M L, et al. Critical applications and prospects of RNA modification in mRNA vaccines (in Chinese). *Chin Sci Bull*, 2024, 69: 4874–4888 [刘聪, 彭金英, 张美玲, 等. RNA修饰在mRNA疫苗中的关键应用和前景. 科学通报, 2024, 69: 4874–4888]

- 24 Vishweshwariah Y L, Dokholyan N V. mRNA vaccines for cancer immunotherapy. *Front Immunol*, 2022, 13: 1029069
- 25 Jalkanen A L, Coleman S J, Wilusz J. Determinants and implications of mRNA poly(A) tail size – Does this protein make my tail look big? *Semin Cell Dev Biol*, 2014, 34: 24–32
- 26 Passmore L A, Coller J. Roles of mRNA poly(A) tails in regulation of eukaryotic gene expression. *Nat Rev Mol Cell Biol*, 2022, 23: 93–106
- 27 Boreikaitė V, Passmore L A. 3'-end processing of eukaryotic mRNA: Machinery, regulation, and impact on gene expression. *Annu Rev Biochem*, 2023, 92: 199–225
- 28 Grier A E, Burleigh S, Sahni J, et al. pEVL: A linear plasmid for generating mRNA IVT templates with extended encoded poly(A) sequences. *Mol Ther Nucleic Acids*, 2016, 5: e306
- 29 Ross J, Sullivan T D. Half-lives of beta and gamma globin messenger RNAs and of protein synthetic capacity in cultured human reticulocytes. *Blood*, 1985, 66: 1149–1154
- 30 Liu X, Huang P, Yang R, et al. mRNA cancer vaccines: Construction and boosting strategies. *ACS Nano*, 2023, 17: 19550–19580
- 31 Karikó K, Muramatsu H, Ludwig J, et al. Generating the optimal mRNA for therapy: HPLC purification eliminates immune activation and improves translation of nucleoside-modified, protein-encoding mRNA. *Nucleic Acids Res*, 2011, 39: e142
- 32 Liu C, Shi Q, Huang X, et al. mRNA-based cancer therapeutics. *Nat Rev Cancer*, 2023, 23: 526–543
- 33 Kong B, Kim Y, Kim E H, et al. mRNA: A promising platform for cancer immunotherapy. *Adv Drug Deliv Rev*, 2023, 199: 114993
- 34 Chaudhary N, Weissman D, Whitehead K A. mRNA vaccines for infectious diseases: Principles, delivery and clinical translation. *Nat Rev Drug Discov*, 2021, 20: 817–838
- 35 Mitchell D A, Batich K A, Gunn M D, et al. Tetanus toxoid and CCL3 improve dendritic cell vaccines in mice and glioblastoma patients. *Nature*, 2015, 519: 366–369
- 36 Deng Z, Tian Y, Song J, et al. mRNA vaccines: The dawn of a new era of cancer immunotherapy. *Front Immunol*, 2022, 13: 887125
- 37 Blass E, Ott P A. Advances in the development of personalized neoantigen-based therapeutic cancer vaccines. *Nat Rev Clin Oncol*, 2021, 18: 215–229
- 38 Hilf N, Kuttruff-Coqui S, Frenzel K, et al. Actively personalized vaccination trial for newly diagnosed glioblastoma. *Nature*, 2019, 565: 240–245
- 39 Shemesh C S, Hsu J C, Hosseini I, et al. Personalized cancer vaccines: Clinical landscape, challenges, and opportunities. *Mol Ther*, 2021, 29: 555–570
- 40 Pan L, Zhang L, Deng W, et al. Spleen-selective co-delivery of mRNA and TLR4 agonist-loaded LNPs for synergistic immunostimulation and Th1 immune responses. *J Control Release*, 2023, 357: 133–148
- 41 Ci L, Hard M, Zhang H, et al. Biodistribution of lipid 5, mRNA, and its translated protein following intravenous administration of mRNA-encapsulated lipid nanoparticles in rats. *Drug Metab Dispos*, 2023, 51: 813–823
- 42 Mockey M, Bourreau E, Chandrashekhar V, et al. mRNA-based cancer vaccine: Prevention of B16 melanoma progression and metastasis by systemic injection of MART1 mRNA histidylated lipopolyplexes. *Cancer Gene Ther*, 2007, 14: 802–814
- 43 Sahin U, Oehm P, Derhovanessian E, et al. An RNA vaccine drives immunity in checkpoint-inhibitor-treated melanoma. *Nature*, 2020, 585: 107–112
- 44 Baden L R, El Sahly H M, Essink B, et al. Efficacy and safety of the mRNA-1273 SARS-CoV-2 vaccine. *N Engl J Med*, 2021, 384: 403–416
- 45 Zhang N N, Li X F, Deng Y Q, et al. A thermostable mRNA vaccine against COVID-19. *Cell*, 2020, 182: 1271–1283.e16
- 46 Alberer M, Gnäd-Vogt U, Hong H S, et al. Safety and immunogenicity of a mRNA rabies vaccine in healthy adults: An open-label, non-randomised, prospective, first-in-human phase 1 clinical trial. *Lancet*, 2017, 390: 1511–1520
- 47 Chen J, Xu Y, Zhou M, et al. Combinatorial design of ionizable lipid nanoparticles for muscle-selective mRNA delivery with minimized off-target effects. *Proc Natl Acad Sci USA*, 2023, 120: e2309472120
- 48 Deng S, Liu Y, Tam R C Y, et al. An intranasal influenza virus-vectored vaccine prevents SARS-CoV-2 replication in respiratory tissues of mice and hamsters. *Nat Commun*, 2023, 14: 2081
- 49 Li M, Li Y, Peng K, et al. Engineering intranasal mRNA vaccines to enhance lymph node trafficking and immune responses. *Acta Biomater*, 2017, 64: 237–248
- 50 van der Ley P A, Zariri A, van Riet E, et al. An intranasal OMV-based vaccine induces high mucosal and systemic protecting immunity against a SARS-CoV-2 infection. *Front Immunol*, 2021, 12: 781280
- 51 Mai Y, Guo J, Zhao Y, et al. Intranasal delivery of cationic liposome-protamine complex mRNA vaccine elicits effective anti-tumor immunity. *Cell Immunol*, 2020, 354: 104143
- 52 Yang J, Zhu J, Sun J, et al. Intratumoral delivered novel circular mRNA encoding cytokines for immune modulation and cancer therapy. *Mol Ther Nucleic Acids*, 2022, 30: 184–197
- 53 Li Q, Ren J, Liu W, et al. CpG oligodeoxynucleotide developed to activate primate immune responses promotes antitumoral effects in combination with a neoantigen-based mRNA cancer vaccine. *Drug Des Devel Ther*, 2021, Volume 15: 3953–3963

- 54 Khazaei Monfared Y, Mahmoudian M, Zakeri-Milani P, et al. Intratumoural delivery of mRNA loaded on a cationic hyper-branched cyclodextrin-based polymer induced an anti-tumour immunological response in melanoma. *Cancers*, 2023, 15: 3748
- 55 Deng Z, Yang H, Tian Y, et al. An OX40L mRNA vaccine inhibits the growth of hepatocellular carcinoma. *Front Oncol*, 2022, 12: 975408
- 56 Qin M, Du G, Sun X. Recent advances in the noninvasive delivery of mRNA. *Acc Chem Res*, 2021, 54: 4262–4271
- 57 Zhao X, Long J, Liang F, et al. Different protective efficacies of a novel antigen-specific DNA vaccine encoding chicken type II collagen via intramuscular, subcutaneous, and intravenous vaccination against experimental rheumatoid arthritis. *Biomed Pharmacother*, 2021, 144: 112294
- 58 Billingsley M M, Singh N, Ravikumar P, et al. Ionizable lipid nanoparticle-mediated mRNA delivery for human CAR T cell engineering. *Nano Lett*, 2020, 20: 1578–1589
- 59 Qin S, Tang X, Chen Y, et al. mRNA-based therapeutics: Powerful and versatile tools to combat diseases. *Sig Transduct Target Ther*, 2022, 7: 166
- 60 Zhang X, Yang B, Ni Q, et al. Materials engineering strategies for cancer vaccine adjuvant development. *Chem Soc Rev*, 2023, 52: 2886–2910
- 61 Zong Y, Wei T, Cheng Q. Recent advances in strategies for developing tissue-selective mRNA-LNP technology (in Chinese). *Chin Sci Bul*, 2024, 69: 4795–4804 [宗岩, 魏妥, 程强. 组织特异性mRNA-LNP递送技术的研发策略. 科学通报, 2024, 69: 4795–4804]
- 62 Wen R, Umeano A C, Kou Y, et al. Nanoparticle systems for cancer vaccine. *Nanomedicine*, 2019, 14: 627–648
- 63 Han X, Zhang H, Butowska K, et al. An ionizable lipid toolbox for RNA delivery. *Nat Commun*, 2021, 12: 7233
- 64 Eygeris Y, Gupta M, Kim J, et al. Chemistry of lipid nanoparticles for RNA delivery. *Acc Chem Res*, 2022, 55: 2–12
- 65 Zhang R, El-Mayta R, Murdoch T J, et al. Helper lipid structure influences protein adsorption and delivery of lipid nanoparticles to spleen and liver. *Biomater Sci*, 2021, 9: 1449–1463
- 66 Ball R L, Hajj K A, Vizelman J, et al. Lipid nanoparticle formulations for enhanced co-delivery of siRNA and mRNA. *Nano Lett*, 2018, 18: 3814–3822
- 67 Yang S T, Kreutzberger A J B, Lee J, et al. The role of cholesterol in membrane fusion. *Chem Phys Lipids*, 2016, 199: 136–143
- 68 Kim J, Eygeris Y, Gupta M, et al. Self-assembled mRNA vaccines. *Adv Drug Deliv Rev*, 2021, 170: 83–112
- 69 Weber J S, Carlino M S, Khattak A, et al. Individualised neoantigen therapy mRNA-4157 (V940) plus pembrolizumab versus pembrolizumab monotherapy in resected melanoma (KEYNOTE-942): A randomised, phase 2b study. *Lancet*, 2024, 403: 632–644
- 70 Liu L, Wang Y, Miao L, et al. Combination immunotherapy of MUC1 mRNA nano-vaccine and CTLA-4 blockade effectively inhibits growth of triple negative breast cancer. *Mol Ther*, 2018, 26: 45–55
- 71 Chen J, Ye Z, Huang C, et al. Lipid nanoparticle-mediated lymph node-targeting delivery of mRNA cancer vaccine elicits robust CD8⁺ T cell response. *Proc Natl Acad Sci USA*, 2022, 119: e2207841119
- 72 Ma S, Li X, Mai Y, et al. Immunotherapeutic treatment of lung cancer and bone metastasis with a mPLA/mRNA tumor vaccine. *Acta BioMater*, 2023, 169: 489–499
- 73 Suberi A, Grun M K, Mao T, et al. Polymer nanoparticles deliver mRNA to the lung for mucosal vaccination. *Sci Transl Med*, 2023, 15: eabq0603
- 74 Tan L, Zheng T, Li M, et al. Optimization of an mRNA vaccine assisted with cyclodextrin-polyethyleneimine conjugates. *Drug Deliv Transl Res*, 2020, 10: 678–689
- 75 Ren J, Cao Y, Li L, et al. Self-assembled polymeric micelle as a novel mRNA delivery carrier. *J Control Release*, 2021, 338: 537–547
- 76 Yang J, Arya S, Lung P, et al. Hybrid nanovaccine for the co-delivery of the mRNA antigen and adjuvant. *Nanoscale*, 2019, 11: 21782–21789
- 77 Cavallaro G, Sardo C, Craparo E F, et al. Polymeric nanoparticles for siRNA delivery: Production and applications. *Int J Pharm*, 2017, 525: 313–333
- 78 Li J, Wu Y, Xiang J, et al. Fluoroalkane modified cationic polymers for personalized mRNA cancer vaccines. *Chem Eng J*, 2023, 456: 140930
- 79 Yin Y, Li X, Ma H, et al. *In situ* transforming RNA nanovaccines from polyethylenimine functionalized graphene oxide hydrogel for durable cancer immunotherapy. *Nano Lett*, 2021, 21: 2224–2231
- 80 Harari A, Graciotti M, Bassani-Sternberg M, et al. Antitumour dendritic cell vaccination in a priming and boosting approach. *Nat Rev Drug Discov*, 2020, 19: 635–652
- 81 Pardi N, Hogan M J, Porter F W, et al. mRNA vaccines — A new era in vaccinology. *Nat Rev Drug Discov*, 2018, 17: 261–279
- 82 Bryant C E, Sutherland S, Kong B, et al. Dendritic cells as cancer therapeutics. *Semin Cell Dev Biol*, 2019, 86: 77–88
- 83 Hosoi A, Takeda Y, Sakuta K, et al. Dendritic cell vaccine with mRNA targeted to the proteasome by polyubiquitination. *Biochem Biophys Res Commun*, 2008, 371: 242–246
- 84 Chung D J, Sharma S, Rangesa M, et al. Langerhans dendritic cell vaccine bearing mRNA-encoded tumor antigens induces antimyeloma immunity after autotransplant. *Blood Adv*, 2022, 6: 1547–1558
- 85 Benteyn D, Heirman C, Bonehill A, et al. mRNA-based dendritic cell vaccines. *Expert Rev Vaccines*, 2015, 14: 161–176
- 86 Higano C S, Small E J, Schellhammer P, et al. Sipuleucel-T. *Nat Rev Drug Discov*, 2010, 9: 513–514
- 87 Kantoff P W, Higano C S, Shore N D, et al. Sipuleucel-T immunotherapy for castration-resistant prostate cancer. *N Engl J Med*, 2010, 363: 411–422

- 88 Di Lorenzo G, Ferro M, Buonerba C. Sipuleucel-T (Provenge®) for castration-resistant prostate cancer. *BJU Int*, 2012, 110: 411
- 89 Perez C R, De Palma M. Engineering dendritic cell vaccines to improve cancer immunotherapy. *Nat Commun*, 2019, 10: 5408
- 90 Scheel B, Teufel R, Probst J, et al. Toll-like receptor-dependent activation of several human blood cell types by protamine-condensed mRNA. *Eur J Immunol*, 2005, 35: 1557–1566
- 91 Liu W, Tang H, Li L, et al. Peptide-based therapeutic cancer vaccine: Current trends in clinical application. *Cell Prolif*, 2021, 54: e13025
- 92 Udhayakumar V K, De Beuckelaer A, McCaffrey J, et al. Arginine-rich peptide-based mRNA nanocomplexes efficiently instigate cytotoxic T cell immunity dependent on the amphipathic organization of the peptide. *Adv Healthcare Mater*, 2017, 6: 1601412
- 93 Pollard C, De Koker S, Saelens X, et al. Challenges and advances towards the rational design of mRNA vaccines. *Trends Mol Med*, 2013, 19: 705–713
- 94 Rosa S S, Prazeres D M F, Azevedo A M, et al. mRNA vaccines manufacturing: Challenges and bottlenecks. *Vaccine*, 2021, 39: 2190–2200
- 95 Huang X, Zhang G, Tang T Y, et al. Personalized pancreatic cancer therapy: From the perspective of mRNA vaccine. *Military Med Res*, 2022, 9: 53
- 96 No authors listed. mRNA vaccine slows melanoma recurrence. *Cancer Discov*, 2023, 13: 1278
- 97 Fotin-Mleczek M, Zanzinger K, Heidenreich R, et al. mRNA-based vaccines synergize with radiation therapy to eradicate established tumors. *Radiat Oncol*, 2014, 9: 180
- 98 Awad M M, Govindan R, Balogh K N, et al. Personalized neoantigen vaccine NEO-PV-01 with chemotherapy and anti-PD-1 as first-line treatment for non-squamous non-small cell lung cancer. *Cancer Cell*, 2022, 40: 1010–1026.e11

Summary for “肿瘤治疗性mRNA疫苗的研发进展”

Advances in the development of therapeutic mRNA vaccines for cancer therapy

Kexin Deng^{1,2†}, Xiaobin Li^{1,2†}, Wanwan Liu^{1,2}, Junyu Liu^{1,2}, Runming Wang^{1,2,3*}, Yandong Yin⁴ & Can Yang Zhang^{1,2,3,4*}

¹ Institute of Biopharmaceutics and Health Engineering, Shenzhen International Graduate School, Tsinghua University, Shenzhen 518055, China

² Key Laboratory of Active Proteins and Peptides Green Biomanufacturing of Guangdong Higher Education Institutes, Tsinghua Shenzhen International Graduate School, Shenzhen 518055, China

³ Key Laboratory of Industrial Biocatalysis, Ministry of Education, Tsinghua University, Beijing 100084, China

⁴ Shenzhen Bay Laboratory, Shenzhen 518032, China

† Equally contributed to this work

* Corresponding authors, E-mail: runmingwang@sz.tsinghua.edu.cn; zhang.cy@sz.tsinghua.edu.cn

In recent years, therapeutic mRNA vaccines for tumors have emerged as a pivotal component of cancer immunotherapy. These vaccines stimulate specific immune responses against tumor-associated or tumor-specific antigens, leveraging the immune system in the body to target and eliminate cancer cells. Composed of mRNA-encoding tumor antigens and delivery vehicles, mRNA vaccines undergo structural modifications to enhance stability and transfection efficiency. Compared to other types of cancer vaccines, mRNA vaccines offer unique advantages. The mRNA vaccines exhibit excellent tolerability *in vivo*, with adverse reactions typically manageable and short-lived. Compared with DNA vaccines, mRNA vaccines do not pose risks of genomic integration and mutations. Moreover, mRNA vaccines avoid the use of pathogenic bacteria or viruses as carriers, ensuring non-infectious profiles. Furthermore, mRNA vaccines can induce sustained and robust humoral and cellular immune responses more effectively than direct antigen injection. The production process of mRNA vaccines is straightforward and cost-effective due to *in vitro* transcription technology. Herein, this review outlines the mechanism of action of therapeutic mRNA vaccines for cancer therapy. Upon administration, mRNA vaccines are internalized by antigen-presenting cells near the injection site. The internalized mRNA is transported to the cytoplasm and ribosomes for translation. This work discusses the commonly targeted antigens in mRNA vaccines, including tumor-associated antigens (TAAs) and tumor-specific antigens (TSAs). TAAs are often overexpressed in tumor cells but possess weaker immunogenicity, which are commonly used in combination to overcome immune tolerance in mammals. TSAs, derived from tumor cell mutations, are highly tumor-specific and immunogenic but elicit weaker tolerance in body. Personalized vaccines targeting unique mutations in cancer patients have been developed clinically, although their cost remains a significant limitation. The review also covers various administration routes for mRNA vaccines, including intravenous, intramuscular, subcutaneous, nasal, and epicutaneous injections. Importantly, delivery systems play a crucial role in mRNA vaccine efficacy due to the molecule's large size and negative charge, which hinder cellular uptake, including lipid-based, polymer-based, dendritic cell-based, and other common carriers, each tailored for specific applications and scenarios. Additionally, lipid nanoparticles (LNP) are currently the most advanced delivery system, employed in approved COVID-19 mRNA vaccines. Finally, this review summarizes current limitations and future challenges facing therapeutic mRNA vaccines for cancer. Addressing these challenges is essential for advancing the development and application of mRNA vaccines in oncology. This comprehensive review not only provides insights into the evolving landscape of therapeutic mRNA vaccines for cancer therapy, but also highlights the mechanisms, advantages, challenges, and potential for cancer combination therapy.

mRNA vaccine, delivery system, administration route, mechanism of action, tumor immunotherapy

doi: [10.1360/TB-2024-0878](https://doi.org/10.1360/TB-2024-0878)