

自噬调控巨噬细胞极化的研究进展

叶莹莹^{1,2}, 康虎², 刘国权^{1,2}, 顾玮^{1,2*}

(¹蚌埠医学院检验医学院, 蚌埠 233030; ²癌症转化医学安徽省重点实验室, 蚌埠 233030)

摘要: 自噬是一种进化上保守的应激反应过程, 将多余的或有潜在危险的胞质成分隔离到双膜小泡自噬小体内, 与溶酶体融合后降解其中产物用于回收利用。巨噬细胞极化是指巨噬细胞在周围环境刺激因子的作用下活化为M1型和M2型, 调节巨噬细胞的活化状态、改善炎症环境是治疗疾病的有效方法。自噬能够改变细胞内代谢状态进而调控巨噬细胞极化。本文将从核因子- κ B(nuclear factor- κ B, NF- κ B)途径、5'AMP激活的蛋白激酶(5'AMP-activated protein kinase, AMPK)/哺乳动物雷帕霉素靶蛋白(mammalian target of rapamycin, mTOR)途径、核苷酸结合寡聚化结构域样受体蛋白-3(nucleotide-binding oligomerization domain-like receptor protein 3, NLRP3)的激活和微小RNA(microRNAs, miRNAs)的调控等方面总结自噬对巨噬细胞活化状态的调控, 为相关疾病治疗提供新策略。

关键词: 自噬; 巨噬细胞极化; 信号通路

Research progress on autophagy regulation of macrophage polarization

YE Yingying^{1,2}, KANG Hu², LIU Guoquan^{1,2}, GU Wei^{1,2*}

(¹School of Laboratory Medicine, Bengbu Medical College, Bengbu 233030, China; ²Key Laboratory of Cancer Translational Medicine of Anhui Province, Bengbu 233030, China)

Abstract: Autophagy is an evolutionarily conservative stress response process that isolates redundant or potentially dangerous cytoplasmic components into the double-membrane vesicle autophagosomes, and degrades the products of recycling after fusion with lysosomes. Macrophage polarization refers to the activation of macrophages into M1 and M2 types under the action of environmental stimulating factors. Regulating the activation state of macrophages to improve the inflammatory environment is effective against treating diseases. Autophagy can change the intracellular metabolic state and regulate the polarization of macrophages. This review will summarize the regulation of autophagy on the activation state of macrophages from the aspects of NF- κ B pathway, AMPK/mTOR pathway, NLRP3 inflammasome activation and regulation of microRNAs, providing new strategies for the treatment of related diseases.

Key Words: autophagy; macrophage polarization; signaling pathways

自噬是细胞中部分胞质元件的自我降解过程, 如错误折叠或聚集的蛋白质、受损的细胞器 (线粒体、内质网和过氧化物酶体) 以及细胞内病原体, 在发育关键时期和应对营养缺乏时, 能平衡

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第一作者: E-mail: yeyingying0709@163.com

*通信作者: E-mail: guwei@bbmc.edu.cn

能量来源^[1]。巨噬细胞是人和动物的天然免疫细胞,广泛存在于体内各个组织中,能维持体内稳态平衡和抵御病原体入侵。巨噬细胞具有可塑性,在局部微环境信号的调节下可被活化为M1型和M2型,分别发挥促炎和抗炎作用,这个过程称为巨噬细胞极化^[2]。有研究发现,自噬在调节巨噬细胞的可塑性中起着至关重要的作用,并参与相应疾病的发生发展过程^[3-6]。本文对近年来有关自噬调控巨噬细胞极化的作用及机制进行总结,以期对相关疾病的研究提供新的靶点。

1 自噬与巨噬细胞极化概述

自噬是一种真核生物进化上保守的核酸和蛋白质降解途径,能清除细胞质成分进而维持细胞内稳态。根据待降解产物递送到溶酶体的方式,可分为三种类型的自噬:巨自噬、微自噬和分子伴侣介导的自噬^[7]。巨自噬是研究自噬的主要范畴,本综述主要关注巨自噬,以下简称为自噬^[8]。目前已鉴定出40个以上的自噬相关基因(*autophagy-related genes*, *ATG*)编码的蛋白质参与自噬不同阶段的调节。在细胞饥饿、缺氧、氧化应激、蛋白质聚集、内质网应激等应激状态下会激活自噬起始UNC-51样激酶-1(*UNC-51-like kinase-1*, *ULK-1*)/*ULK-2*复合物,该复合物磷酸化Ⅲ类PI3K(*phosphatidylinositol-3-kinase*, *PI3K*)复合物,在自噬前体膜上产生磷脂酰肌醇-3-磷酸,促进吞噬体成核^[9,10]。磷脂酰肌醇-3-磷酸和*ATG5-ATG12-ATG16L1*复合物等促进磷脂酰乙醇胺与微管相关蛋白-1-轻链-3(*microtubule-associated protein light chain 3*, *LC3*)结合,生成*LC3 II*引导吞噬体膜延伸闭合为自噬小体^[11]。随后,在可溶性N-乙基马来酰亚胺敏感因子附着蛋白受体的介导下,自噬小体与溶酶体融合后降解其所包裹的内容物,以实现细胞本身的代谢需要和细胞器的更新^[12]。

人体内的巨噬细胞主要来源于骨髓和胚胎时期的卵黄囊、胎儿肝脏^[13]。巨噬细胞极化是指巨噬细胞响应微环境刺激和信号而极化成不同的表型,可激活为M1型(经典激活)和M2型(替代激活)^[14]。脂多糖和Th1型细胞因子如干扰素- γ 和肿瘤坏死因子- α ,能诱导巨噬细胞M1型极化,其特征是上调共刺激分子CD80、CD86、诱导型一氧化氮

合酶(*inducible nitric oxide synthase*, *iNOS*)以及各种促炎细胞因子和趋化因子的表达,包括肿瘤坏死因子- α 、白介素-1 α (*interleukin-1 α* , *IL-1 α*)、*IL-1 β* 、*IL-6*、*IL-12*等。*IL-4*、*IL-13*、*IL-10*和转化生长因子- β 可诱导巨噬细胞M2型极化,主要表现为甘露糖受体CD206、膜蛋白CD163以及多种抗炎细胞因子和趋化因子,如*IL-10*、转化生长因子- β 等表达增加^[14]。M1型和M2型巨噬细胞的功能不同,M1型巨噬细胞参与炎症促进、趋化作用、自由基形成、基质降解、抗菌和抗肿瘤效应;而M2型巨噬细胞有利于炎症消解、寄生虫清除、血管生成和组织修复;同时,M2型巨噬细胞还可抑制效应性T细胞介导的免疫反应,促进组织重塑,促进肿瘤发展^[14,15]。

巨噬细胞作为先天免疫系统的重要组成部分,参与防御病原体入侵、清除细胞碎片和调节炎症反应,组织炎症的程度在很大程度上取决于巨噬细胞活化后的表型^[15]。自噬也与巨噬细胞中包括巨噬细胞极化在内的许多功能密切相关^[8]。自噬相关基因*ATG5*敲除小鼠会出现肝脏和全身炎症,巨噬细胞向M1型极化,导致肝损伤的进展^[16]。自噬缺陷的乳腺癌肺转移细胞分泌的巨噬细胞移动抑制因子,诱导巨噬细胞向M1型极化也证实了这一点^[17]。Li等^[18]报道,海带多糖培养小鼠巨噬细胞后,巨噬细胞自噬蛋白的表达增加,且向M2型极化;而自噬抑制剂3-甲基腺嘌呤处理后,能阻断海带多糖对巨噬细胞极化的影响。因此,自噬蛋白表达或自噬通量的降低或升高均会影响巨噬细胞的极化方向。

2 自噬调控巨噬细胞极化的相关分子机制

2.1 核因子NF- κ B参与自噬调控巨噬细胞极化

核因子- κ B(*nuclear factor- κ B*, *NF- κ B*)调节巨噬细胞中许多重要基因的表达,还参与单核细胞的招募并使其分化为巨噬细胞^[19],靶向NF- κ B是自噬调控巨噬细胞极化的重要机制^[20,21]。NF- κ B是由Rel蛋白家族的同源或异源二聚体组成的转录因子,即p65(Rel A)、p50(NF- κ B1)、p52(NF- κ B2)、c-Rel和Rel B。活化的p50/p65异源二聚体在M1极化后可促进*iNOS*、*IL-12*等炎症相关基因的表达,p50/p50同源二聚体能与其竞争炎症相关基因启动

子的结合部位, 阻断基因的转录^[20]。Taetzsch等^[22]也发现, NF- κ B p50是调节小胶质细胞M1/M2平衡的关键信号。Li等^[23]报道, 小鼠巨噬细胞与 β -葡聚糖共培养后, 自噬蛋白LC3 II蛋白表达降低, M1型巨噬细胞标志物iNOS、CD80 mRNA表达增加, NF- κ B p65的蛋白质表达增加; 而NF- κ B抑制剂SN50可逆转 β -葡聚糖诱导的巨噬细胞M1型极化和自噬蛋白表达降低。因此, β -葡聚糖可通过NF- κ B p65依赖的自噬途径将巨噬细胞极化为M1型, 会增加动脉粥样硬化的风险。自噬也能降低肿瘤相关巨噬细胞中NF- κ B p65的稳定性, 用肝癌细胞的上清液培养骨髓来源的巨噬细胞后, 细胞表面Toll样受体-2(Toll-like receptor 2, TLR2)会通过自噬途径下调NF- κ B p65蛋白表达, 促进巨噬细胞向M2型极化^[24]。肝细胞癌患者中高水平的高迁移率族蛋白-1与疾病的严重程度相关, 该蛋白质使巨噬细胞LC3 II蛋白表达增加, NF- κ B p65蛋白表达减少, 促使巨噬细胞向M2型极化^[25]。自噬可影响NF- κ B p65的表达水平, 改变巨噬细胞的极化方向, NF- κ B p65上调导致巨噬细胞向M1型极化, NF- κ B p65下调使巨噬细胞向M2型极化^[23,25,26]。

2.2 AMPK/mTOR途径参与自噬调控巨噬细胞极化

5'AMP激活的蛋白激酶(5'AMP-activated protein kinase, AMPK)是一种保守的三磷酸腺苷敏感酶, 还是氧化应激感受器和氧化还原调节的靶点^[27]。mTOR是自噬的负调控因子, 在饥饿状态下, mTORC1对ULK1/ULK2复合物的抑制作用解除; AMPK可负向调节mTOR, 也能磷酸化ULK1/ULK2复合物启动自噬^[9]。AMPK/mTOR信号通路与巨噬细胞极化有关, 二甲双胍和膜联蛋白A1经AMPK/mTOR通路诱导巨噬细胞向M2型转化, 可促进血管生成, 防止脑缺血损伤^[28,29]。生物素A促进巨噬细胞M1型极化, 也通过AMPK/ULK1/mTOR通路介导自噬, 抵御沙门氏菌感染^[30]。用乳腺癌细胞4T1上清液培养小鼠巨噬细胞后, 其LC3 II蛋白表达减少且向M2型极化; 加入mTOR的自噬诱导剂雷帕霉素后, 巨噬细胞向M1型极化^[31]。5-氨基乙酰丙酸介导的非致死性声动力疗法通过活性氧(reactive oxygen species, ROS)/AMPK/mTORC1/自噬途径, 促进巨噬细胞向M2型极化,

增强胆固醇外流和抗炎反应, 增加动脉粥样硬化斑块的稳定性, 预防血栓形成^[32]。AMPK/mTOR通路参与自噬对巨噬细胞极化的调控, 但其中详细的调控机制尚不完全清楚, 需进一步研究。

2.3 NLRP3炎性体参与自噬调控巨噬细胞极化

核苷酸结合寡聚化结构域样受体蛋白-3(nucleotide-binding oligomerization domainlike receptor protein 3, NLRP3)是一种胞内多蛋白复合体, 可在病原体 and 损伤相关模式分子的共同作用下诱导炎症和嗜热细胞死亡^[33]。NLRP3炎性体的激活可诱导天冬氨酸特异性半胱氨酸蛋白酶-1(Caspase-1)和IL-1 β 的前体切割, 变为成熟形式并将其释放到下游的级联免疫反应中^[29]。自噬可清除NLRP3炎性体激活剂, 如细胞内的阻滞剂、NLRP3炎性体成分和细胞因子, 减轻炎症反应^[6,33]。NLRP3炎性体具有促炎作用, 与巨噬细胞的M1型极化有关, 在大鼠牙根吸收模型中, NLRP3炎性体激活, 诱导巨噬细胞M1型极化^[34]。研究表明, 自噬可通过激活NLRP3炎性体进而调控巨噬细胞极化^[35,36]。山奈酚可促进自噬, 抑制NLRP3炎性体活化和巨噬细胞M1型极化, 减轻角膜移植中的排斥反应^[35]。去泛素化酶19(ubiquitin-specific protease 19, USP19)可增加自噬通量, 减少线粒体活性氧的产生, 抑制NLRP3炎性体的激活, 促进人源巨噬细胞向M2型极化^[36]。促进自噬可抑制NLRP3炎性体的激活进而改变巨噬细胞的极化方向, 将NLRP3炎性体的促炎功能转变为抗炎功能, 为炎症性疾病提供新的治疗靶点, 但其分子机制仍需进一步研究。

2.4 微小RNA参与自噬对巨噬细胞极化的调控

微小RNA(microRNAs, miRNAs)是一种长度为20~22个核苷酸的非编码RNA分子, 主要功能是通过与靶标mRNA的3'非翻译区结合, 转录后阻止mRNA翻译或启动mRNA降解^[37]。MiRNAs参与自噬的不同阶段, 包括自噬诱导、自噬小体成核、自噬小体延伸、自噬小体与溶酶体融合, 还调节自噬诱导的上游信号通路^[38,39]。MiR-9、miR-127、miR-155和miR-125b促进巨噬细胞M1型极化, miR-124、miR-223、miR-34a、let7c、miR-132、miR-146a和miR-125a-5p通过靶向多种转录因子和接头蛋白诱导巨噬细胞M2型极化^[40]。

MiRNAs还可同时调控自噬与巨噬细胞极化, miR-498通过鼠双微体蛋白-2抑制转录激活因子-3降解, 抑制食管癌中巨噬细胞的自噬和M2型极化^[41]; miR-375抑制巨噬细胞极化调控转录因子Kruppel样因子-4(kruppel-like factor-4, KLF4)的激活, 可参与巨噬细胞的自噬抑制和M1型极化^[42]。在IL-6/pSTAT/miR-155-3p/自噬/pSTAT3正反馈环中, miR-155-3p激活多形性胶质母细胞瘤中的信号转导与转录激活因子-3(signal transducer and activator of transcription 3, STAT3)启动自噬进而诱导巨噬细胞M2型极化, 促进胶质瘤的进展^[43,44]。Jiang等^[45]发现, 脂肪干细胞来源的外泌体中miR-30d-5p表达增加, 能抑制自噬介导的小胶质细胞向M1型转化, 进而预防脑损伤。综上, miRNAs可分别调控自噬和巨噬细胞极化, 也可通过调控自噬过程介导巨噬细胞极化。

2.5 其他途径参与自噬对巨噬细胞极化的调控

除NF- κ B、AMPK/mTOR、NLRP3炎症体和miRNAs途径外, 自噬和巨噬细胞极化还有其他潜在的联系。二十二碳六烯酸增强巨噬细胞KLF4的表达, 磷酸化p38丝裂原活化蛋白激酶(mitogen activated protein kinase, MAPK), 上调自噬蛋白LC3 II表达, 促使巨噬细胞向M2型极化^[46]。肿瘤细胞释放的自噬小体(tumor cell-released autophagosomes, TRAPS)经TLR4介导的髓样分化因子-88(myeloid differentiation factor 88, MyD88)/p38/STAT3途径可诱导巨噬细胞向M2型极化^[47]。钛(Ti)种植体、载银TiO₂纳米管材料也参与自噬调控巨噬细胞的极化^[48,49]。电化学阳极氧化法制备的载银纳米TiO₂纳米管(Ag@TiO₂-NTs)经表面修饰后能控制释放超低剂量Ag⁺离子, 抑制PI3K/蛋白激酶B(protein kinase B, Akt)途径及下游葡萄糖转运蛋白-1的表达, 激活自噬, 诱导巨噬细胞向M2型转化^[48]。外力刺激牙周膜干细胞后, 自噬蛋白LC3 II表达增加, Akt磷酸化被抑制; 受刺激后的牙周膜干细胞上清液与人源巨噬细胞共培养后, 后者向M1型极化^[50]。自噬是在应激、炎症、低氧和机械负荷等外界刺激下维持细胞和组织动态平衡的过程, 其上下游均受多种信号机制调控。以上研究表明, 自噬可调控巨噬细胞极化, 影响相关疾病的发生发展和转归。

3 自噬调控巨噬细胞极化在相关疾病中的作用

巨噬细胞极化存在于癌症、动脉粥样硬化、类风湿关节炎等多种疾病过程中, 且在疾病发生发展中起重要作用^[2]。自噬也与许多疾病的进程有关, 包括免疫性疾病、癌症、神经退行性疾病、心血管疾病和衰老^[8]。目前有研究表明, 一些药物可通过自噬调控巨噬极化发挥作用^[3-5]。木皂苷C通过沉默调节蛋白-1(silent information regulator 1, Sirt1)介导的自噬影响巨噬细胞极化, 减少泡沫样细胞的形成, 从而减轻动脉粥样硬化^[3]。在乳腺癌肿瘤微环境模型中, 自噬可通过ROS/胞外信号调节激酶(extracellular signal-regulated kinase, ERK)和AMPK/mTOR信号通路, 抑制异丙肾上腺素诱导的M2型极化^[30]。黄秋葵总黄酮靶向自噬介导巨噬细胞极化, 进而重塑肠道微生物群, 抑制慢性肾功能衰竭产生的微炎症^[51]。曲古菌素A(trichostatin A, TSA)通过增强自噬、促进巨噬细胞M2型极化, 减少全身炎症, 可提高多菌败血症小鼠的存活率^[52]。中药复方消水汤(xiaoshui decoction, XSD)在体外实验中可促进自噬, 使巨噬细胞向M1型极化, 可用于恶性胸腔积液的治疗^[5]。在不同的疾病中, 促进自噬并不完全指向巨噬细胞M1型极化。富氢盐水抑制肺泡巨噬细胞自噬可驱使巨噬细胞向M2型转化, 减轻脂多糖诱导的急性肺损伤; 自噬抑制剂3-甲基腺嘌呤抑制肺泡巨噬细胞自噬, 巨噬细胞M2型极化增加也从侧面证实了这一点^[53]。由此可见, 自噬可改变巨噬细胞的极化状态, 产生促炎或抗炎效果, 最终影响疾病发生发展的方向(表1)。

4 总结与展望

自噬作为一种应激条件下的生存机制, 不仅维持细胞内稳态, 还能影响炎症和免疫系统调节, 这些功能都与巨噬细胞极化有关。自噬在巨噬细胞极化调控过程中发挥着重要的作用, NF- κ B途径、AMPK/mTOR途径、NLRP3炎症体以及miRNAs直接或间接参与了自噬对巨噬细胞极化的调控。其他疾病中的研究也证实, 在动脉粥样硬化、败血症、恶性胸腔积液等疾病中, BS-KS、

表1 自噬调控巨噬细胞极化的相关分子机制

涉及因素	自噬水平	作用机制	巨噬细胞极化方向	相应疾病	参考文献
CD5L	↑	上调转录因子ID3	M2	感染、动脉粥样硬化、癌症	[54]
木皂苷C	↑	上调Sirt1	M2	动脉粥样硬化	[3]
USP19	↑	抑制NLRP3炎性体激活	M2	炎症反应	[36]
异丙肾上腺素	↓	ROS/ERK和mTOR途径	M2	乳腺癌	[31]
二十二碳六烯酸	↑	p38MAPK途径	M2	慢性炎症疾病	[46]
黄秋葵总黄酮	↓	AMPK/Sirt1途径	M2	慢性肾衰	[51]
MiR-30d-5p	↓	抑制自噬	M1	急性缺血性卒中	[45]
TSA	↑	促进自噬	M2	脓毒症	[52]
链脲佐菌素(STZ)	↓	溶酶体功能障碍	M1	糖尿病并发症	[55]
补肾抗衰老片	↑	PPAR- γ /NF- κ B途径	M2	动脉粥样硬化	[4]
精胺(SPM)	↑	上调ATG5	M2	急性肝损伤	[56]
β -葡聚糖	↓	上调NF- κ Bp65	M1	动脉粥样硬化	[23]
Ag@TiO ₂ -NTs	↑	PI3K/Akt途径	M2	骨愈合	[48]
XSD	↑	促进自噬	M1	恶性胸腔积液	[5]
TRAPs	↑	MyD88/p38/STAT3途径	M2	肿瘤	[47]
钛(Ti)种植体	↑	ERK/Beclin1途径	M2	组织愈合	[49]
外力刺激	↑	Akt/NF- κ B途径	M1	炎症性骨重塑	[50]
MiR-155-3p	↑	IL-6/pSTAT3/miR-155-3p/自噬/STAT3途径	M2	胶质瘤	[44]
山奈酚	↑	抑制NLRP3炎性体激活	M2	角膜移植排斥反应	[35]
丹参酮II A	↑	miR-375/KLF4途径	M2	动脉粥样硬化	[42]
富氢盐水(HRS)	↓	抑制自噬	M2	急性肺损伤	[53]
非致死性声动力疗法	↑	ROS/AMPK/mTORC1途径	M2	动脉粥样硬化	[32]
海带多糖	↑	增强自噬	M2	动脉粥样硬化	[18]
TLR2信号	↑	下调NF- κ Bp65	M2	肝癌	[24]

注：自噬水平中箭头↑表示自噬相关蛋白表达增加或自噬通量增加，箭头↓则相反

TSA、XSD作用后自噬蛋白表达或自噬通量发生改变，巨噬细胞极化状态也随之改变，提示自噬可介导巨噬细胞的极化，进而影响疾病进程，其相关基因成为疾病潜在的生物学标志和新的治疗靶点。但到目前为止，关于自噬调控巨噬细胞极化状态的研究还存在许多问题，自噬基因是否直接调节巨噬细胞极化的信号通路，自噬是否调节巨噬细胞极化关键蛋白的降解等还需要进一步研究。随着对自噬以及巨噬细胞极化调控机制的深入认识，我们对相关疾病的防治水平也将有所提高。

参考文献

- [1] Glick D, Barth S, Macleod KF. Autophagy: cellular and molecular mechanisms. *J Pathol*, 2010, 221(1): 3-12
- [2] Shapouri-Moghaddam A, Mohammadian S, Vazini H, et al. Macrophage plasticity, polarization, and function in health and disease. *J Cell Physiol*, 2018, 233(9): 6425-6440
- [3] Luo Y, Lu S, Gao Y, et al. Araloside C attenuates atherosclerosis by modulating macrophage polarization via Sirt1-mediated autophagy. *Aging (Albany NY)*, 2020, 12(2): 1704-1724
- [4] Xie Y, Tian L, Fang Z, et al. Bushen kangshuai tablet inhibits progression of atherosclerosis by intervening in macrophage autophagy and polarization. *J Tradit Chin Med*, 2020, 40(1): 28-37
- [5] Jin Z, Shen C, Zhang H, et al. Chinese medicine xiaoshui decoction inhibits malignant pleural effusion in mice and mediates tumor-associated macrophage polarization by activating autophagy. *J Ethnopharmacol*, 2020, 249: 112412
- [6] Yu SX, Du CT, Chen W, et al. Genipin inhibits NLRP3 and NLRC4 inflammasome activation via autophagy

- suppression. *Sci Rep*, 2016, 5(1): 17935
- [7] Boya P, Reggiori F, Codogno P. Emerging regulation and functions of autophagy. *Nat Cell Biol*, 2013, 15(7): 713-720
- [8] Wu MY, Lu JH. Autophagy and macrophage functions: inflammatory response and phagocytosis. *Cells*, 2019, 9(1): 70
- [9] Mizushima N, Levine B. Autophagy in human diseases. *N Engl J Med*, 2020, 383(16): 1564-1576
- [10] Chang C, Jensen LE, Hurley JH. Autophagosome biogenesis comes out of the black box. *Nat Cell Biol*, 2021, 23(5): 450-456
- [11] Choi AMK, Ryter SW, Levine B. Autophagy in human health and disease. *N Engl J Med*, 2013, 368(7): 651-662
- [12] Lamark T, Svenning S, Johansen T. Regulation of selective autophagy: the p62/SQSTM1 paradigm. *Essays Biochem*, 2017, 61(6): 609-624
- [13] Chen X, Tang J, Shuai W, et al. Macrophage polarization and its role in the pathogenesis of acute lung injury/acute respiratory distress syndrome. *Inflamm Res*, 2020, 69(9): 883-895
- [14] Yunna C, Mengru H, Lei W, et al. Macrophage M1/M2 polarization. *Eur J Pharmacol*, 2020, 877: 173090
- [15] Sica A, Erreni M, Allavena P, et al. Macrophage polarization in pathology. *Cell Mol Life Sci*, 2015, 72(21): 4111-4126
- [16] Liu K, Zhao E, Ilyas G, et al. Impaired macrophage autophagy increases the immune response in obese mice by promoting proinflammatory macrophage polarization. *Autophagy*, 2015, 11(2): 271-284
- [17] Cotzomi-Ortega I, Nieto-Yañez O, Juárez-Avelar I, et al. Autophagy inhibition in breast cancer cells induces ROS-mediated MIF expression and M1 macrophage polarization. *Cell Signal*, 2021, 86: 110075
- [18] Li XY, Wang YJ, Chen S, et al. Laminaria japonica polysaccharide suppresses atherosclerosis via regulating autophagy-mediated macrophage polarization. *J Agric Food Chem*, 2022, 70(12): 3633-3643
- [19] Juhas U, Ryba-Stanisawska M, Szargiej P, et al. Different pathways of macrophage activation and polarization. *Postepy Hig Med Dosw (Online)*, 2015, 69: 496-502
- [20] Kuo WT, Chang JM, Chen CC, et al. Autophagy drives plasticity and functional polarization of tumor-associated macrophages. *IUBMB Life*, 2022, 74(2): 157-169
- [21] Biswas SK, Lewis CE. NF- κ B as a central regulator of macrophage function in tumors. *J Leukocyte Biol*, 2010, 88(5): 877-884
- [22] Taetzsch T, Levesque S, McGraw C, et al. Redox regulation of NF- κ B p50 and M1 polarization in microglia. *Glia*, 2015, 63(3): 423-440
- [23] Li X, Luo H, Ye Y, et al. Betaglucon, a dectin1 ligand, promotes macrophage M1 polarization via NF- κ B/autophagy pathway. *Int J Oncol*, 2019, 54(1): 271-282
- [24] Chang CP, Su YC, Hu CW, et al. TLR2-dependent selective autophagy regulates NF- κ B lysosomal degradation in hepatoma-derived M2 macrophage differentiation. *Cell Death Differ*, 2013, 20(3): 515-523
- [25] Shiau DJ, Kuo WT, Davuluri GVN, et al. Hepatocellular carcinoma-derived high mobility group box 1 triggers M2 macrophage polarization via a TLR2/NOX2/autophagy axis. *Sci Rep*, 2020, 10(1): 13582
- [26] Huang YJ, Hung KC, Hung HS, et al. Modulation of macrophage phenotype by biodegradable polyurethane nanoparticles: possible relation between macrophage polarization and immune response of nanoparticles. *ACS Appl Mater Interfaces*, 2018, 10(23): 19436-19448
- [27] Wang Y, Liu Z, Shu S, et al. AMPK/mTOR signaling in autophagy regulation during cisplatin-induced acute kidney injury. *Front Physiol*, 2020, 11: 619730
- [28] Xu X, Gao W, Li L, et al. Annexin A1 protects against cerebral ischemia-reperfusion injury by modulating microglia/macrophage polarization via FPR2/ALX-dependent AMPK-mTOR pathway. *J Neuroinflammation*, 2021, 18(1): 119
- [29] Qing L, Fu J, Wu P, et al. Metformin induces the M2 macrophage polarization to accelerate the wound healing via regulating AMPK/mTOR/NLRP3 inflammasome signaling pathway. *Am J Transl Res*, 2019, 11(2): 655-668
- [30] Zhao X, Tang X, Guo N, et al. Biochanin A enhances the defense against salmonella enterica infection through AMPK/ULK1/mTOR-mediated autophagy and extracellular traps and reversing SPI-1-dependent macrophage (M Φ) M2 polarization. *Front Cell Infect Microbiol*, 2018, 8: 318
- [31] Shan M, Qin J, Jin F, et al. Autophagy suppresses isoprenaline-induced M2 macrophage polarization via the ROS/ERK and mTOR signaling pathway. *Free Radical Biol Med*, 2017, 110: 432-443
- [32] Yang Y, Wang J, Guo S, et al. Non-lethal sonodynamic therapy facilitates the M1-to-M2 transition in advanced atherosclerotic plaques via activating the ROS-AMPK-mTORC1-autophagy pathway. *Redox Biol*, 2020, 32: 101501
- [33] Biasizzo M, Kopitar-Jerala N. Interplay between NLRP3 inflammasome and autophagy. *Front Immunol*, 2020, 11: 591803
- [34] Zhang J, Liu X, Wan C, et al. NLRP3 inflammasome mediates M1 macrophage polarization and IL-1 β production in inflammatory root resorption. *J Clin Periodontol*,

- 2020, 47(4): 451-460
- [35] Tian H, Lin S, Wu J, et al. Kaempferol alleviates corneal transplantation rejection by inhibiting NLRP3 inflammasome activation and macrophage M1 polarization via promoting autophagy. *Exp Eye Res*, 2021, 208: 108627
- [36] Liu T, Wang L, Liang P, et al. USP19 suppresses inflammation and promotes M2-like macrophage polarization by manipulating NLRP3 function via autophagy. *Cell Mol Immunol*, 2021, 18(10): 2431-2442
- [37] Wang S, Huang Y, Zhou C, et al. The role of autophagy and related microRNAs in inflammatory bowel disease. *Gastroenterol Res Pract*, 2018, 2018: 1-10
- [38] Akkoc Y, Gozuacik D. MicroRNAs as major regulators of the autophagy pathway. *Biochim Biophys Acta Mol Cell Res*, 2020, 1867(5): 118662
- [39] Yao H, Han B, Zhang Y, et al. Non-coding RNAs and autophagy. *Adv Exp Med Biol*, 2019, 1206: 199-220
- [40] Essandoh K, Li Y, Huo J, et al. MiRNA-mediated macrophage polarization and its potential role in the regulation of inflammatory response. *Shock*, 2016, 46(2): 122-131
- [41] Li D, Yan M, Sun F, et al. MiR-498 inhibits autophagy and M2-like polarization of tumor-associated macrophages in esophageal cancer via MDM2/ATF3. *Epigenomics*, 2021, 13(13): 1013-1030
- [42] Chen W, Li X, Guo S, et al. Tanshinone IIA harmonizes the crosstalk of autophagy and polarization in macrophages via miR-375/KLF4 pathway to attenuate atherosclerosis. *Int Immunopharmacol*, 2019, 70: 486-497
- [43] Xue H, Yuan G, Guo X, et al. A novel tumor-promoting mechanism of IL6 and the therapeutic efficacy of tocilizumab: Hypoxia-induced IL6 is a potent autophagy initiator in glioblastoma via the p-STAT3- MIR155-3p-CREBRF pathway. *Autophagy*, 2016, 12(7): 1129-1152
- [44] Xu J, Zhang J, Zhang Z, et al. Hypoxic glioma-derived exosomes promote M2-like macrophage polarization by enhancing autophagy induction. *Cell Death Dis*, 2021, 12(4): 373
- [45] Jiang M, Wang H, Jin M, et al. Exosomes from miR-30d-5p-ADSCs reverse acute ischemic stroke-induced, autophagy-mediated brain injury by promoting M2 microglial/macrophage polarization. *Cell Physiol Biochem*, 2018, 47(2): 864-878
- [46] Kawano A, Ariyoshi W, Yoshioka Y, et al. Docosahexaenoic acid enhances M2 macrophage polarization via the p38 signaling pathway and autophagy. *J Cell Biochem*, 2019, 120(8): 12604-12617
- [47] Wen ZF, Liu H, Gao R, et al. Tumor cell-released autophagosomes (TRAPs) promote immunosuppression through induction of M2-like macrophages with increased expression of PD-L1. *J Immunother cancer*, 2018, 6(1): 151
- [48] Chen Y, Guan M, Ren R, et al. Improved immunoregulation of ultra-low-dose silver nanoparticle-loaded TiO₂ nanotubes via M2 macrophage polarization by regulating GLUT1 and autophagy. *Int J Nanomedicine*, 2020, 15: 2011-2026
- [49] Luo J, He Y, Meng F, et al. The role of autophagy in M2 polarization of macrophages induced by micro/nano topography. *Int J Nanomedicine*, 2020, 15: 7763-7774
- [50] Jiang N, He D, Ma Y, et al. Force-induced autophagy in periodontal ligament stem cells modulates M1 macrophage polarization via AKT signaling. *Front Cell Dev Biol*, 2021, 9: 666631
- [51] Tu Y, Fang QJ, Sun W, et al. Total flavones of *abelmoschus manihot* remodels gut microbiota and inhibits microinflammation in chronic renal failure progression by targeting autophagy-mediated macrophage polarization. *Front Pharmacol*, 2020, 11: 566611
- [52] Cui SN, Chen ZY, Yang XB, et al. Trichostatin A modulates the macrophage phenotype by enhancing autophagy to reduce inflammation during polymicrobial sepsis. *Int Immunopharmacol*, 2019, 77: 105973
- [53] Qiu P, Liu Y, Chen K, et al. Hydrogen-rich saline regulates the polarization and apoptosis of alveolar macrophages and attenuates lung injury via suppression of autophagy in septic rats. *Ann Transl Med*, 2021, 9(12): 974
- [54] Sanjurjo L, Aran G, Téllez É, et al. CD5L promotes M2 macrophage polarization through autophagy-mediated upregulation of ID3. *Front Immunol*, 2018, 9: 480
- [55] Yuan Y, Chen Y, Peng T, et al. Mitochondrial ROS-induced lysosomal dysfunction impairs autophagic flux and contributes to M1 macrophage polarization in a diabetic condition. *Clin Sci(Lond)*, 2019, 133(15): 1759-1777
- [56] Zhou S, Gu J, Liu R, et al. Spermine alleviates acute liver injury by inhibiting liver-resident macrophage pro-inflammatory response through ATG5-dependent autophagy. *Front Immunol*, 2018, 9: 948